

Modelling Microbial Fuel Cells using lattice Boltzmann methods

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Abstract

An accurate modelling of bio-electrochemical processes that govern Microbial Fuel Cells (MFCs) and mapping their behaviour according to several parameters will enhance the development of MFC technology and enable their successful implementation in well defined applications. The geometry of the electrodes is among key parameters determining efficiency of MFCs due to the formation of a biofilm of anodophilic bacteria on the anode electrode, which is a decisive factor for the functionality of the device. We simulate the bio-electrochemical processes in an MFC while taking into account the geometry of the electrodes. Namely, lattice Boltzmann methods are used to simulate the fluid dynamics and the advection–diffusion phenomena in the anode compartment. The model is verified on voltage and current outputs of a single MFC derived from laboratory experiments under continuous flow. Conclusions can be obtained from a parametric analysis of the model concerning the design of the geometry of the anode compartment, the positioning and microstructure of the anode electrode, in order to achieve more efficient overall performance of the system. An example of such a parametric analysis is presented here, taking into account the positioning of the electrode in the anode compartment.

Keywords: microbial fuel cells, lattice Boltzmann, modelling, agent-based model, bio-electrochemical processes

Introduction

Due to improvements in power and current outputs achieved in the last decades [1], studies in Microbial Fuel Cells (MFCs) have intensified [2]. Despite the increased interest in this field, the majority of research work focuses on practical experimental analysis. On the other hand, just a handful of studies [3] have steered towards mathematically describing and simulating the bio-electrochemical processes that generate the electrical output of these devices. Arguably, the intrinsic complexity of these systems requires highly elaborative procedures to produce accurate models. However, the advantages (in time and cost saving terms) acquired by an accurate MFC model has motivated researchers [4, 5, 6] to exert more effort towards this direction.

The utilisation of porous electrodes in continuous flow MFCs, which is realised as best practice due to higher outputs [7], and the observation that the macrostructure of the anode, its material and its inhomogeneities have a significant impact [8, 9], prompted the employment of the lattice Boltzmann method (LBM) in the framework of the MFC model presented here. LBM is a numerical tool based on mesoscopic phenomena and is able to model fluid dynamics and transport (advection–diffusion) phenomena in fluid systems [10]. In contrast to its precursor, lattice gas automata that studies the individual molecular dynamics of fluids or gases –in a microscopic scale–, LBM simulates transport phenomena by studying the evolution of

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ensembles of molecules –in a mesoscopic scale. Nonetheless, the basic principles of microscopic physics are retained due to the utilisation of the ensemble averaged distribution function as the primary variable [10]. LBMs have an inherent parallel nature that is advantageous when implementing the algorithm in specialised parallel hardware. Moreover, the lattices used can trivially illustrate complex geometries compared with other computation methods.

The ultimate goal of the proposed model is to simulate in an efficient way the behaviour of an MFC device, namely the voltage and current output. That is achieved by analysing in the macroscopic and mesoscopic scales several phenomena occurring in the anode half-cell of an MFC –assuming that the cathode performance is not limiting. Specifically, fluid dynamics, transport phenomena, bio-electrochemical processes and biofilm formation are all taken into consideration and, thus, simulated by separate procedures, that constitute the proposed model whilst interacting with each other. The formation of a biofilm of anodophilic bacteria on the anode electrode is a decisive factor for the functionality of the device. Fluid dynamics and transport phenomena were simulated using two different LBMs, one providing the velocity of the fluid throughout the porous electrode positioned in the anode compartment and the other calculating the concentration of chemicals –given the velocity field acquired from the first LBM.

The use of LBMs, in addition to providing the ability to represent inhomogeneities or complex geometries of electrodes, empowers the faster execution of critical calculations that can be considered as the bottleneck of previous studies that have studied these phenomena [11]. That is due to the numerous iterations needed for the system to reach an equilibrium in the procedures of fluid dynamics and transport phenomena and the frequency at which these two procedures have to be updated, namely every time the biofilm formation is updated.

To support the aforementioned statement, note that the deciding chemical, physical and biological processes occurring in an MFC need to be studied across a wide span of time scales [12, 13]. Although the time for a system to reach an equilibrium concerning its fluid dynamics and transport of chemicals, is in the period of some seconds, the evolution –or even a small change of the formation– of a biofilm is noted after some hours or days. As a result, when modelling the evolution in the behaviour of a system that includes such a complicated collection of interdisciplinary processes, the faster relaxing (reaching equilibrium) procedures, here fluid dynamics and advection-diffusion, will have to be revised every time the slower updating process provides slightly different outputs. That introduces a bottleneck in the efficient execution of such detailed models.

The proposed model can provide a parametric analysis concerning the design of the geometry of the anode compartment, the positioning and microstructure of the anode electrode. The geometry of the anode compartment and the positioning of the electrode have a significant role as they can ensure a sufficient nutrient delivery to the entire biofilm surface. Moreover, the microstructure of the anode electrode can enhance an efficient attachment and growth of the biofilm. The aforementioned aspects are decisive factors in reaching higher power production from the biofilm.

1. Previous work

LBMs have been extensively studied and claimed to be a viable alternative to other computational methods [10]. This method can be used for different modelling purposes, such as reaction-diffusion systems, complex fluid dynamics or wave propagation [10, 14]. LBM is considering a mesoscale statistical representation of groups of molecules and is divided into two processes, the collision and the propagation of these groups. The collision operator characterising each LBM, leads to the relaxation of the distributions of the groups to a local equilibrium which is the result of a set of parameters like the velocity, the concentration (considered as conserved quantities) and external forces.

The robustness of different types and variations of LBMs have been studied for several problems. For instance, in [15], the fluid dynamics through complex geometries, namely porous media, was evaluated for several types of LBM and with different boundary conditions. In [16], a three-dimensional LBM was suggested, implementing twenty seven characteristic velocities to model the flow field in porous media. The authors paralleled the results from their model with the ones from other numerical models and measurements from experiments.

In addition to fluid dynamics, LBM has successfully tackled the solution of advection–diffusion equations with high accuracy [17]. Moreover, the ability of LBM to model nonlinear convection–diffusion phenomena with anisotropic diffusion have been presented in [18], where the modelling of several one- and two-dimensional paradigms was investigated in detail. In [19] a comparison of different lattices has been executed to analyse the robustness and accuracy of them. Also, the impact that the chosen scheme of boundary conditions has in the accuracy of the model was studied [20].

Moreover, researchers have previously employed the LBM in the field of conventional fuel cells. For instance, in [21] the diffusion of gases –that are characterised by different molecular weights– were modelled in complex porous media, specifically in a solid oxide fuel cell (SOFC) anode. The authors in [22] studied the flow field in a three-dimensional scheme of a section of a serpentine channel and in a two-dimensional scheme of a channel filled or partially filled with a porous medium, taking particular interest in the boundary conditions and the simulated time periods. Moreover, in [23] a multi-component flow model was presented to analyse the transport of a mixture of gases in a SOFC anode through irregular porous media. The impact of the micro-structure of the electrode, the dimensionless reaction flux and fuel composition to the fuel cell outputs were studied. Also, the effect that the micro-structure of the anode has on the distribution and deposition of carbon and, thus, to the performance of the cell was studied [24].

In addition, LBMs have been used in three-dimensional simulation of biofilm growth and evolution with respect to the flow dynamics and the advection–diffusion of chemicals through porous media [12, 25]. Two types of biofilms were studied, with zero permeability [12] and non-zero permeability [25]. The main difference in the work presented here with the aforementioned, is that here the function of the anode compartment of an MFC is simulated, namely, the bio-electrochemical reactions were studied in addition to the flow dynamics, transport phenomena and biofilm formation. Nonetheless, the impact that the bio-electrochemical phenomena have towards all the other procedures were included to the model. Keeping in mind the ultimate goal of an enhanced performance of the model, the presented model is based on a unified discrete grid where all procedures of the model are calculated. On the other hand, previous works [12, 25] have used LBMs based on discrete grids, whereas the simulation of biofilm is tackled with an individual-based model [26] utilising continuous Cartesian dimensions. This incoherency in the procedures of the model will pose difficulties while implementing the model efficiently in software (or indeed hardware).

In the present work the selection of using a two-dimensional grid, rather than a three-dimensional similar to previous studies, certainly produces a lower accuracy, as claimed in [12, 25]. However, the use of a two-dimensional grid enables a faster execution of the model, whilst scaling up the grid to three dimensions is trivial. Moreover, the utilisation of the same grid for all the procedures of the model, is an additional advantage when advancing towards three dimensions.

As mentioned before, a great advantage of LBMs is their capacity of significant acceleration which is straightforward, due to the inherently parallel characteristics of LBM. The parallel implementation of the algorithm of the model can be realised with advanced programming languages compatible with multi-core processors [27] or specialised hardware –like Field Programmable Gate Arrays (FPGAs) [28, 29] or Graphics Processing Units (GPUs) [30].

The biofilm formation is a process that it is not yet fully decoded by scientific research. The accurate modelling of biofilms is a quite demanding task, given that the behaviour of living, self-organising organisms has to be taken into consideration and the experimental justification of simulated results is challenging. However, several examples of previous work aimed to explain and model the underlying mechanics of the evolution of the biofilm, like attachment, growth, decay, lysis and detachment of biomass [13, 26, 31, 32]. A simple agent-based model that can simulate the basic procedures of attachment and development was used here. Note here, that despite the simplicity of the model, it is capable of robustly formulating the under study processes.

Further advantages of the LBM model are that a detailed analysis of mass transfer can be carried out for the actual micro-structure of the anode electrode and the cell’s performance can be judged by calculating the concentration polarisation. Complex geometries can be easily handled and both continuum and non-continuum diffusion through the pores can be modelled. Because of the level of detail, the effect of anode geometry on voltage polarisation can be directly studied and the geometry can be optimised to provide enhanced performance. In fact, being able to model an anodic micro-structure may be advantageous in

understanding 3D porous electrode behaviour of real MFCs and their biofilms.

2. Methods

To approximate the behaviour of a MFC the model simulates the formation of a biofilm on the electrode surface, the fluid dynamics and the advection–diffusion phenomena throughout the anode compartment and calculates outputs of the MFC based on local concentration of chemicals and electrochemical equations. Two LBMs were used. One LBM represents the dynamics of fluids inside the anode compartment and another LBM represents advection-diffusion equation of chemicals, specifically the carbon source for the biofilm in the feedstock (acetate in this study). The model runs as follows:

1. Formation of the electrode geometry and initialisation of the parameters.
2. Calculation of fluid dynamics with LBM (first LBM model).
3. Calculation of the advection–diffusion equation with LBM (second LBM model).
4. Calculation of MFC outputs (bio-electrochemical model).
5. Biofilm formation.
6. Go to step (2).

2.1. Lattice Boltzmann Method

A LBM is characterised by three main factors: the lattice, the equilibrium equation and the kinetic equation. We adopt the Bhatnagar-Gross-Krook (BGK) LBM framework [33] with a D2Q9 lattice (Fig. 1), based on a two-dimensional lattice with nine characteristic velocities:

$$c_0 = (0, 0) \tag{1}$$

$$c_{1,3} = (\pm 1, 0) \quad c_{2,4} = (0, \pm 1) \tag{2}$$

$$c_{5,6,7,8} = (\pm 1, \pm 1) \tag{3}$$

The particle velocities distribution is governed by the evolution equation which takes into account propagation and collision processes:

$$F_i(x + c_i \Delta t, t + \Delta t) - F_i(x, t) = -\frac{\Delta t}{\tau} \left[F_i(x, t) - F_i^{(0)}(x, t) \right] \tag{4}$$

where τ is the dimensionless relaxation time, t is a simulation time step, x is a site in the lattice, and i is an index of the characteristic velocity. $F_i^{(0)}(x, t)$ is the equilibrium distribution function:

$$F_i^{(0)}(\rho, j) = \frac{W_i}{\rho_0} \rho \left[1 + 3 \frac{c_i \cdot u}{c^2} + \frac{9}{2} \frac{(c_i \cdot u)^2}{c^4} - \frac{3}{2} \frac{u^2}{c^2} \right] \tag{5}$$

where c is a model constant relating to the lattice speed ($c = \frac{dx}{dt \cdot \sqrt{3}}$), c_i are defined in Eqs. (1-3), u is the velocity flow, ρ is the mass density and $\frac{W_i}{\rho_0}$ is a weight factor equal to:

$$\frac{W_i}{\rho_0} = \frac{4}{9}, \text{ for } i = 0 \tag{6}$$

$$\frac{W_i}{\rho_0} = \frac{1}{9}, \text{ for } i = 1, 2, 3, 4 \tag{7}$$

$$\frac{W_i}{\rho_0} = \frac{1}{36}, \text{ for } i = 5, 6, 7, 8. \tag{8}$$

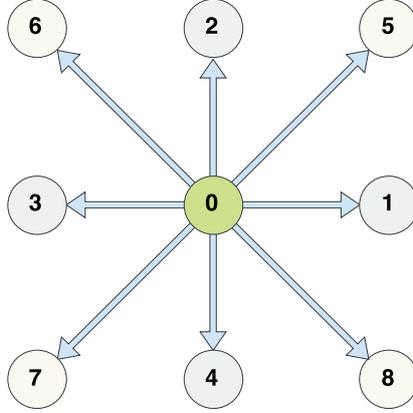


Figure 1: The D2Q9 lattice.

2.1.1. Fluid dynamics

Fluid dynamics are calculated by LBM with boundaries defined by the compartment physical boundaries (i.e. plastic container), the anode electrode geometry and the dynamic biofilm formation. The method is based on the BGK model, see Eqs. (1 – 8). This method is utilising bounce-back boundary conditions [10] and has a second order accuracy for fluid dynamics. The resulting mass density (ρ) and momentum density (j) determine the flow field as follows:

$$\rho(x, t) = \sum_i F_i(x, t) \quad (9)$$

$$j(x, t) = \rho(x, t)u(x, t) = \sum_i c_i F_i(x, t) \quad (10)$$

On the boundaries the bounce-back principle dictates that the evolution equation is not calculated as in Eq. (4). However, the collision process is updated depending on the configuration of the boundaries and the available space. For instance, in the case of a boundary placed in the south direction of the lattice (Fig. 2) the following equations apply:

$$F_5(x, t) = F_7(x, t), \quad F_2(x, t) = F_4(x, t), \quad F_6(x, t) = F_8(x, t). \quad (11)$$

2.1.2. Advection–diffusion equations

The advection–diffusion equations are solved using another BGK LBM, where the flow field and geometry of electrodes are used as inputs. The boundary conditions are considered as Dirichlet boundaries for the input and output of the domain (with a given concentration amount) and as Neumann for the solid boundaries (with a set zero-flux of concentration towards them) [34]. The evolution equation for the distribution function is similar to Eq. (4) (note the different notation of the distribution function used – G_i rather than F_i – to distinguish the two separate LBMs):

$$G_i(x + c_i \Delta t, t + \Delta t) - G_i(x, t) = -\frac{\Delta t}{\tau_D} \left[G_i(x, t) - G_i^{(0)}(x, t) \right] \quad (12)$$

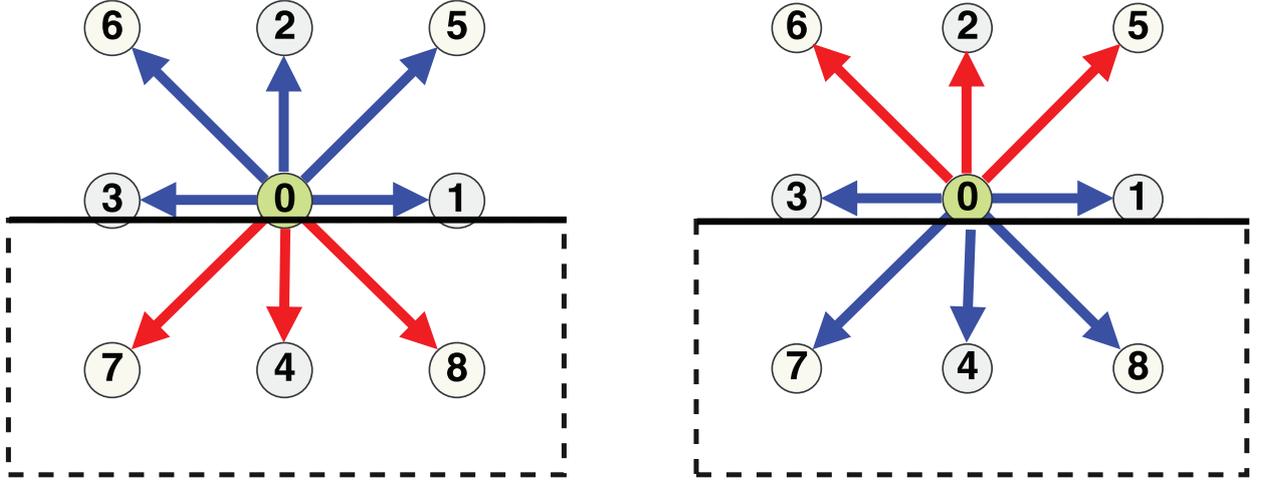


Figure 2: The bounce-back boundaries principle in a D2Q9 lattice.

where $G_i^{(0)}(x, t)$ is the equilibrium distribution function, which is calculated as in Eq. (5) and W_i are the weights defined by Eqs. (6)-(8). Note that the velocity (u) is considered as a constant for the advection-diffusion simulation and provided by the first LBM procedure.

The resulting local concentration (C_s) which represents the concentration field is calculated in the following :

$$C_s(x, t) = \sum_i G_i(x, t) \quad (13)$$

2.2. Bio-electrochemical model

The mechanism for the transfer of electrons to the anode is assumed to incorporate the existence of an intracellular mediator and the interconnection of the biomass with the electrode by nanowires or by direct contact [5, 35]. In the simulated experiments the total concentration of the intracellular mediator is regarded as constant and analogous to the biomass concentration. After calculating the equilibrium of fluid dynamics and the concentration of the chemical species, the output of MFC is studied using the following equations:

$$M_{total} = M_{red} + M_{ox} \quad (14)$$

$$M_{ox}(x, t + 1) = M_{ox}(x, t) - Yq_a + \gamma \frac{I_{cell}}{mF} \frac{1}{V_a C^{bio}} \quad (15)$$

where M_{total} is the total amount of intracellular mediator, M_{red} is the reduced form, while M_{ox} is the oxidised form. Y is the mediator yield granted by the utilisation of the substrate, γ is the mediator molar mass, m is the fraction of electrons provided during reduction of the intracellular mediator. F is the Faraday constant, V_a is the anode compartment volume, C^{bio} is the concentration of biomass. I_{cell} is the total current produced and, thus, flowing through the device and q_a is the consumption rate of the substrate.

The double Monod equation is used to determine the consumption rate of the substrate:

$$q_a = q_{max} \frac{C_s}{C_s + K_s} \frac{M_{ox}}{M_{ox} + K_{M_{ox}}} \quad (16)$$

where C_s is the concentration of the substrate, K_s is the half saturation constant for substrate, $K_{M_{ox}}$ is the half saturation constant for oxidised mediator and q_{max} is the maximum consumption rate.

The MFC as an electrochemical system suffers from losses, defined elsewhere as polarisation or overpotential, caused by the rates of the reactions (activation overpotential), the ohmic resistances of the material

used to build the MFC (ohmic overpotential) and the limitations posed on the mass transfer or kinetic phenomena (concentration overpotential). As a result, considering Kirchhoff's voltage law and Ohm's law the following equation correlates the overpotentials, the values of resistance and the produced current.

$$I_{cell}R_{ext} = E_0 - I_{cell}R_{int} - n_{conc} - n_{act} \quad (17)$$

where R_{ext} and R_{int} are the external (load) and the internal resistances, E_0 is the maximum produced voltage (or the open circuit voltage), n_{conc} and n_{act} are the concentration and activation overpotentials, respectively. The concentration overpotential based on the Nerst equation is defined by:

$$n_{conc} = \frac{RT}{F} \ln \left(\frac{M_{total}}{M_{red}} \right) \quad (18)$$

where R is the universal gas constant and T is the temperature; while, the activation overpotential can be calculated by an approximation of the Butler-Volmer equation:

$$n_{act} = \frac{I_{cell}}{A_a I_0} \frac{RT}{mF} \left(\frac{M_{ox}}{M_{red}} \right) \quad (19)$$

where A_a is the anode electrode surface area and I_0 is the exchange current density for mediator oxidation in reference conditions. Consequently, the output current can be estimated by:

$$I_{cell} = \frac{(E_0 - n_{conc} - n_{act})}{(R_{int} + R_{ext})} \frac{M_{red}}{\varepsilon + M_{red}} \quad (20)$$

where ε is a constant which bounds the output current at low values of M_{red} . While, Ohm's law provides the voltage produced:

$$V_{cell} = I_{cell}R_{tot} = I_{cell} \left(R_{int} + R_{ext} \right) \quad (21)$$

2.3. Biofilm formation

The biofilm formation is an agent-based procedure simulating the attachment and growth of biomass. The attachment of biomass is simulated by a random selection in each time step of a predefined number of lattice cells (k_{ata}) to change their biomass concentration to a predefined initial value ($C^{bio} = C_0^{bio}$). The random selection of these cells is amongst the ones representing the front between the fluid domain and the anode electrode domain. Note that the initial value of the intracellular mediator in the cells selected for biomass attachment is set and equivalent to the biomass predefined initial value.

The growth of the biomass is following the double Monod equation providing the consumption rate (Eq. (16)). Along with the growth of biomass, an analogous increase of the intracellular mediator is assumed. The spreading of the biomass is initiated when the biomass concentration in a lattice cell exceeds a predefined threshold (C_{max}^{bio}). The direction towards the spreading of the biomass happens, is randomly selected. If the under study cell is surrounded by cells representing unavailable space (solid boundaries and existing amounts of biomass) a random walk is executed to find a free cell towards where a displacement of biomass can occur. When a lattice cell representing an available choice is found, each cell with an existing value of biomass, transports its biomass value to the neighbour indicated by the random walk. Finally, when an adjacent cell of the initial cell exceeding the predefined threshold is freed up, a fraction of the biomass concentration of the initial cell (fr_{spr}) is subtracted from its biomass concentration and it is transported to the currently free cell. In addition to the transport of biomass being displaced by the random walk, the amounts of intracellular mediators follow the same procedure.

Note here that the initial densities of attached biomass -forming the biofilm- are considered as in previous studies of anodophilic biofilm [13, 35]. The biofilm is initially formed in fronts between bulk fluid domain and the anode electrode and that is the reason for modelling the biomass density initially in that domain. The initial formation of the biofilm is realized during the inoculation process of the experimental procedure,

which is not studied by the model presented here. Different environmental conditions and input parameters can be adopted during the inoculation process that will affect the initial formation of the biofilm, hence, the parameter C_0^{bio} of the simulation. Despite this fact, the latter parameter is considered as described previously, since it proves to have a negligible impact on the final results; it mainly affects how fast the biofilm is formed initially because then the growth procedure is more significant.

The initial concentration of biomass attached to the electrode is a result of the inoculation procedure, while during the continuous flow operation of an MFC more biomass can attach on the anode electrode. One of the assumptions of the model was that the biofilm is the primary producer of electrons, thus current. As a result, the anodophilic bacteria that are present in the bulk space of the anode compartment are not studied in terms of growth, producing electrons or their detailed movement around the anode compartment. Finally, as mentioned before, the mechanisms that dictate the formation and growth of biofilm are not perfectly known and as a result difficult to validate in experiments. Thus, the model uses a stochastic process when determining the procedure of the biofilm evolution; however the double Monod equation was used to determine the rate of biomass growth.

The biofilm will be accumulating, for as long as there is acetate around and available space in the anode compartment; however, this is not entirely accurate. This is definitely a limitation of the present version of the model; nonetheless, further work is required to align the model with the realistic behaviour of biofilms, especially, in mixed species communities as the ones accommodated in MFCs and study its effects in the performance of these systems.

Following the completion of the biofilm formation procedure, all the algorithmic steps of the model are repeated, starting with the calculation of fluid dynamics with LBM as formerly defined. The fluid flow is recalculated taking into account the updated biofilm as impermeable. Consequently, the advection and diffusion of the chemicals have to also be updated. Then, the electrical outputs are revised and, finally, the biofilm with the new concentrations and flow field is reformed. The model was implemented with Matlab R2016b.

3. Results

The applicability of the model can be verified with the comparison of the simulated results with measurements obtained in the laboratory. More specifically, here, the measurements obtained by an experiment with an innovative configuration of a single MFC, located onto a MFC system based on a building brick, were used. The MFC is comprised by the anode compartment accommodated in one cavity of the brick, the cathode compartment in another cavity and the ceramic semi-permeable separator between them. The dimensions of the anode compartment, which are the same as the cathode, are depicted in Table 1, while the separator thickness is 18.5 mm. Anode electrodes were made of 20 g/m² carbon fibre veil (PRF Composites, UK) with dimensions as given in Table 1. The MFC was inoculated with a mixture of activated sewage sludge (Cam Valley, Wessex Water Scientific Laboratory, Saltford, UK) and fresh urine and operated in continuous mode [36, 37]. The output of the MFC connected with a 360 Ω resistor as an electrical load was measured at 300 mV and 830 μA.

The influent of the anode compartment is considered for the simulation and depicted in the following figures to enter from the left side of the x -axis. Consequently, the effluent is considered to exit the anode compartment from the right side of the x -axis of the illustrated model configuration.

The values of the parameters used in the aforementioned equations that describe the functionality of the MFC are given in Table 1. These values are extracted by measurements in the laboratory experiment or adopted by previous attempts of describing MFCs and biofilms with mathematical equations in the literature [13, 35].

The procedures of calculating fluid dynamics and the advection–diffusion equation are running for enough time steps until an equilibrium is reached, while the bio-electrochemical model and the procedure of biofilm formation are updated once during each simulation interval. One iteration of the model corresponds to one hour of real time.

In Fig. 3 the simulated anode compartment configuration is depicted. The black lines on top and on the bottom of the figure represents the impermeable boundaries (chassis) of the anode compartment. The

Parameter	Description	Value
$X \times Y \times Z$	Anode compartment dimensions	$17 \times 60 \times 65mm$
$X' \times Y' \times Z'$	Anode electrode dimensions	$17 \times 48 \times 65mm$
$L_X \times L_Y$	Model lattice dimensions	60×65
ϕ	Electrode porosity	0.874
V	Fluid input velocity	$1.758 \cdot 10^{-2}mm/s$
ν	Kinematic viscosity	$1.004mm^2/s$
τ	Dimensionless relaxation time (LBM)	0.6706
D	Diffusion coefficient	$0.0012mm^2/s$
τ_D	Dimensionless relaxation time (LBM)	0.5036
C_s^{in}	Input substrate concentration	$410 mg substrate / L$
$Mtotal$	Total amount of intracellular mediator	$0.05 \frac{mg mediator}{mg biomass}$
Y	Intracellular mediator yield	$0.5687 \frac{mg mediator}{mg substrate}$
γ	Mediator molar mass	$663400 \frac{mg mediator}{mol mediator}$
m	Electrons provided during reduction	$2 e$
q_{max}	Maximum consumption rate	$8.48 \frac{mg substrate}{mg biomass \cdot day}$
K_S	Half saturation constant for substrate	$20 (mg substrate / L)$
K_{Mox}	Half saturation constant for oxidised mediator	$0.02 \times Mtotal (mg mediator/L)$
I_0	Exchange current density	$0.001 A/m^2$
E_0	Open circuit voltage	$0.7 V$
R_{ext}	External ohmic resistance (load)	360Ω
R_{int}	Internal ohmic resistance	360Ω
k_{ata}	Predefined number of cells for attachment	200
C_0^{bio}	Initial biomass concentration	$450mg biomass / L$
C_{max}^{bio}	Threshold biomass concentration	$512.5 mg biomass / L$
f_{rspr}	Fraction of the biomass spreading	40%

Table 1: Parameters used in the execution of the model.

influent is entering the compartment from $x = 0$ and the effluent is exiting from $x = 60$. Whereas, the porous medium of the anode electrode is illustrated with the black shapes that are repeated throughout the figure. Note that a random geometry of the anode electrode was adopted. Nonetheless, this selection imposes no limitation on the functionality of the model. The formation of the geometry of the anode electrode can be disorganised, lacking a regular geometry or order, or based on mathematical self-organising mathematical tools [38].

The flow and concentration field in the anode compartment can be calculated without any biofilm formed on the anode electrode. The results of such calculations are illustrated in Figs. 4 and 5.

When a biofilm is developed on the anode electrode (illustrated in Fig. 6 as the gray spots attached on the black shapes that represent the porous electrode) then the flow and concentration fields are recalculated taking into consideration the new configuration. The results of the flow field LBM calculation are depicted in Fig. 7 and the results of the advection-diffusion LBM are depicted in Fig. 8.

Finally, the simulation of the functionality of the under study MFC is carried out for a simulation of 72 hours and the results are presented in the following figures. In Fig. 9 the output voltage of the MFC is illustrated. The output voltage is asymptotically increasing during the first 60 hours and then reaches a steady state of approximately 330 mV that is in agreement with the laboratory based measurements. Moreover, the output current depicted in Fig. 10 is following the increase of the voltage curve and it reaches a steady value of 0.91 mA (similar with the laboratory experiment).

In Fig. 11 the ratio of the reduced and the oxidised mediator to the total amount of intracellular mediator are depicted through time. The concentration of reduced mediator (solid line) is following the evolution of the output voltage, whereas the concentration of the oxidised mediator (dashed line) is following a complete opposite asymptotic curve of the reduced mediator, as derived by Eq. (14). Note here that the model is

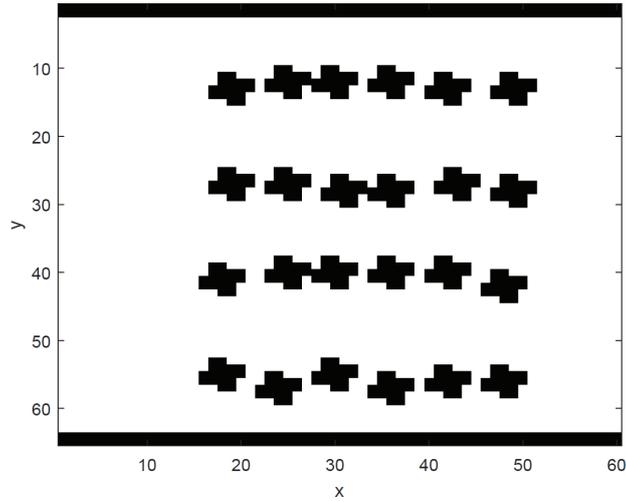


Figure 3: The geometry of the porous anode electrode.

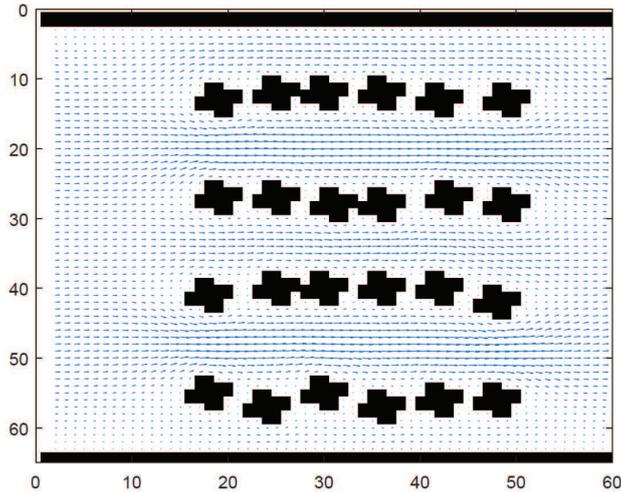


Figure 4: Flow field in the anode without a developed biofilm.

studying intracellular mediators' concentrations. As mentioned before, an assumption is made that they are analogous to the biomass concentration and for abstraction reasons this analogy is considered constant. This explains the equilibrium between the reduced and the oxidized mediator presented in Fig. 11.

Nonetheless, in order to study the effect of the positioning of the anode electrode and the sensitivity of the outputs produced by the model to the inputs depicting the details of a system, different electrode geometries were used as inputs. The first geometry (depicted in Fig. 3) is simulating an electrode that is filling the whole of the anode compartment, with the granules representing its microstructure placed with random distances among them. The second geometry tested (see Fig. 12) is also simulating an electrode filling the whole of the anode compartment, but with a more canonical microstructure (granules are placed in a fixed grid). Moreover, testing the placement of the electrode in a partially filled anode, the third geometry (Fig. 13) and fourth geometry (Fig. 14) were used.

The comparison of the results provided by the model is illustrated in Fig. 15. It is clear that even the slightest difference in the microstructure of the anode electrode (like the small deviation among electrode granules between first -Fig. 3- and second geometry -Fig. 12) has a distinct effect in the voltage output of

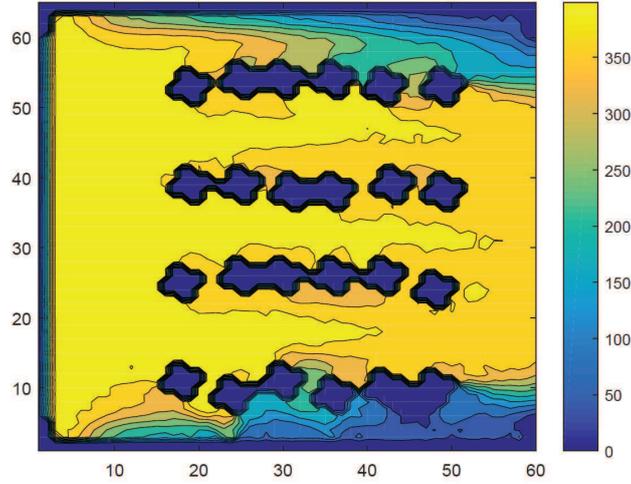


Figure 5: Concentration field of acetate in the feedstock in the anode without a developed biofilm (in mg/L).

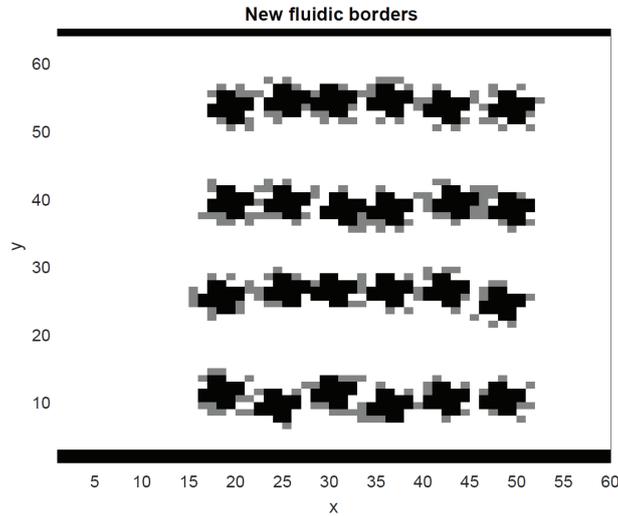


Figure 6: The configuration of the initial biofilm on the anode electrode porous medium.

the system. The electrode of the first geometry gives a higher voltage faster; a fact that can be attributed to the slightly longer distances between the granules placed near the inlet of the system, thus, acetate reaches a greater area of the biofilm faster. Nonetheless, the third and fourth geometries have higher voltage outputs compared with the first and second, because of the same reason as in the previous. Finally, the geometry with the two sections of the electrodes placed in the sides of the compartment is the most efficient from the ones studied, as the specific geometry facilitates the development of the biofilm and, at the same time, a higher amount of acetate is reaching the region occupied by biofilm.

No other parameter was changed in the four cases, other than the positioning and microstructure of the anode electrode, and this is probably the reason why the resultant voltages are similar (although the first and second geometries in time step representing the 72th hour do show a difference of 2.5% (at 0.3335V) and 5% (at 0.3257V), respectively).

The formation of the biofilm around the anode electrode is considered as a very important factor of the final performance of a MFC [4, 8, 11]. Here, the formation of the biofilm is simulated, as previously described, by an algorithm that depends on the maximum amount of biomass that can be present in a

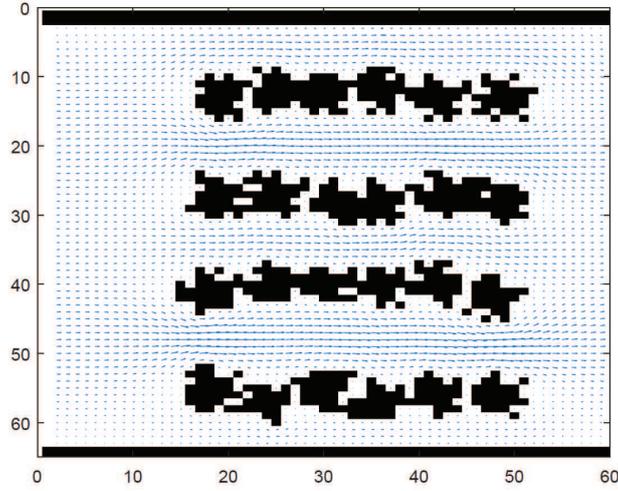


Figure 7: Flow field in the anode after the development of the initial biofilm.

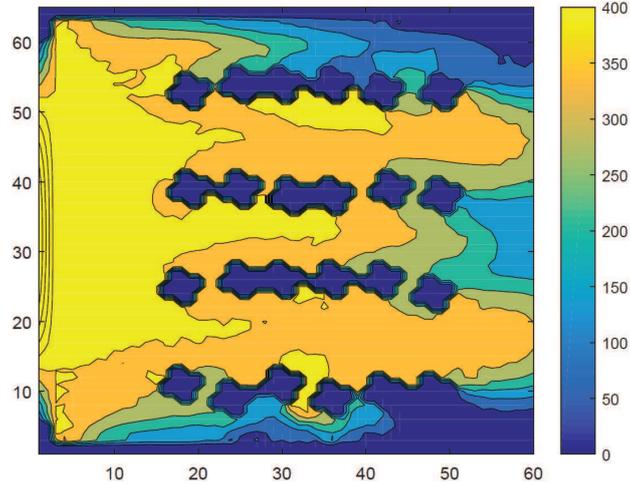


Figure 8: Concentration field of acetate in the feedstock in the anode after the development of the initial biofilm (in mg/L).

simulated area (C_{max}^{bio}) and the local growth rate of biofilm. The biofilm modeled here, is not essentially of constant depth around the anode boundaries, because of the random initialization, attachment and the different local concentrations of nutrients that trigger an uneven growth. In a real MFC inside which the biofilm is formed from pure electroactive species, the depth (or thickness) will be uniform, however this requires further simulation work to demonstrate this in a model. Finally, the simulated biofilm will be comprised of CA cells that have biomass concentrations between 60% and 100% of the C_{max}^{bio} parameter.

Despite the fact that only the steady state outputs of the MFCs studied here, were reported, the time dependence of the outputs (Fig. 15) follow the norm established by experimental and simulated outputs described in previous studies [4, 5, 8, 11]. Voltage and current are building up as a concave function to reach a plateau defined by the steady state outputs of the aforementioned experiments [36, 37] within some days.

4. Conclusions

We proposed a model of MFC that incorporate geometry of the electrodes. The model implements two LBMs to calculate the fluid flow field and the advection–diffusion phenomena of chemicals inside the

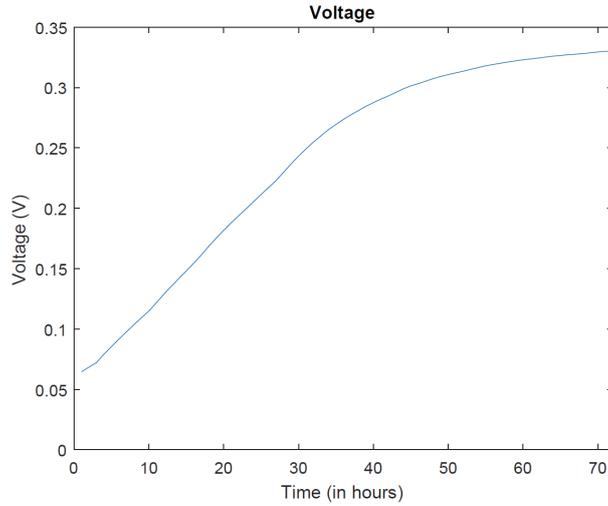


Figure 9: MFC output voltage over time in hourly intervals.

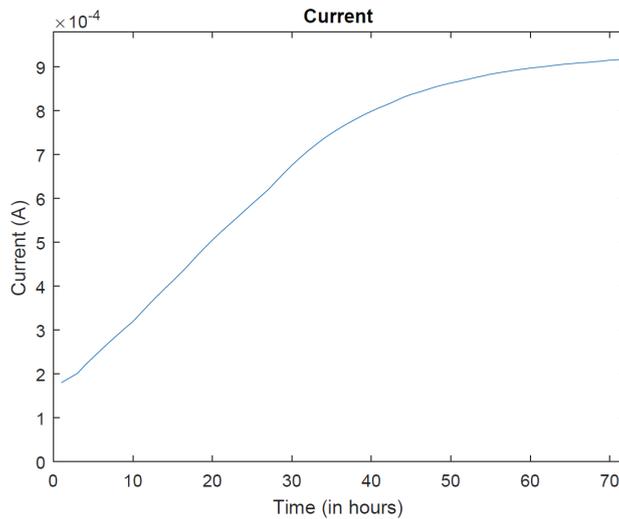


Figure 10: MFC output current over time in hourly intervals.

anode compartment. Also, an agent-based model calculates the attachment and spreading of biomass on the electrode. The output voltage is calculated using bio-electrochemical processes.

An advantage of the model is that it is designed to consider the flow dynamics of the influent and the advection-diffusion phenomena of chemicals in comparably short time intervals (seconds and minutes). Thus, the model can be utilised in predicting transitions of the behaviour of a MFC from one state to another in this time span when the relevant parameter, namely influent velocity or fuel concentration is altered.

We adopted a random geometry of the anode electrode. This imposes no limitations on the model's functionality. The formation of the geometry of the anode electrode can be disorganised, lacking a regular geometry or order, or based on mathematical self-organising mathematical tools [38]. Studying particular geometries will be a topic of future research. This could include analysis of the effect of specified geometries and micro-structures on the development of biofilms in a MFC and its electrical and chemical outputs, e.g. geometries based data from X-ray tomographic microscopy [16].

The biofilm formed during modelled development of a MFC is assumed to impose a very high resistance

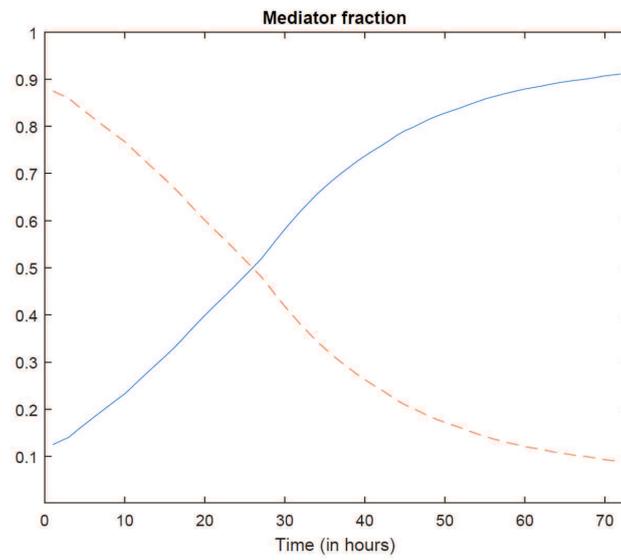


Figure 11: Intracellular mediator concentrations fractions over time in hourly intervals (reduced – solid line, oxidised – dashed line).

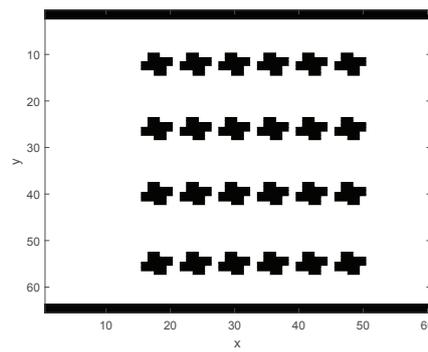


Figure 12: The second test geometry of the porous anode electrode.

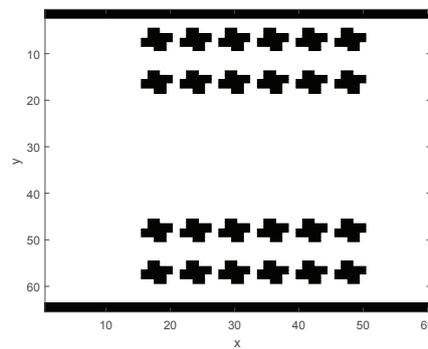


Figure 13: The third test geometry of the porous anode electrode.

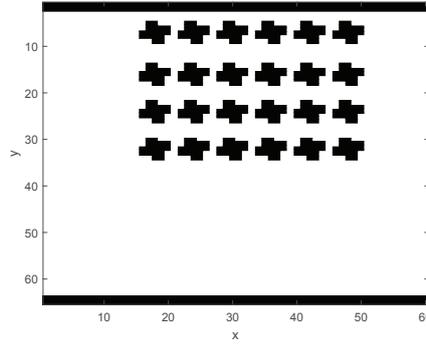


Figure 14: The fourth test geometry of the porous anode electrode.

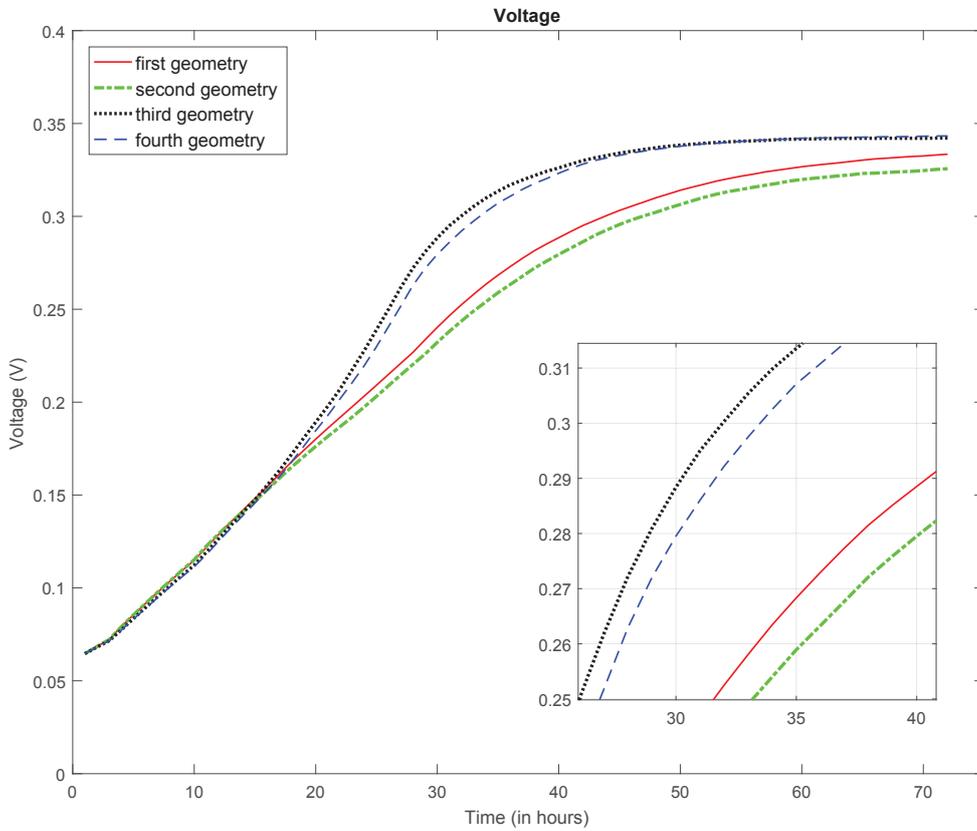


Figure 15: Comparison of simulated voltage outputs for different anode electrode.

to fluid dynamics, thus, the speed of fluid components throughout the biofilm was considered negligible. While in the area that biofilm is present there is zero velocity assumed, as described previously, the diffusion phenomena occur without any change on the diffusivity. However, this is not an ideal representation of a real biofilm. More work would be required to take into account the permeability of biofilms and communication between cells in the matrix.

While, in the simulated experiments the total concentration of the intracellular mediator is regarded as constant and analogous to the biomass concentration, in reality, fractions of oxidised and reduced forms of

the mediator can vary. This could be another aspect of future work. Finally, the parametric analysis of positioning and microstructure of the geometry of the anode electrode in lab experiments will be further investigated. As a result, we anticipate establishing, if possible, a more direct connection between the outputs of the model and the real-life system.

Acknowledgment

This work was supported by the European Union’s Horizon 2020 Research and Innovation Programme under Grant Agreement No. 686585.

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