

1 **Title:** Potential for molecular mimicry between the human endogenous retrovirus W family envelope  
2 proteins and myelin proteins in multiple sclerosis.

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11 **Running Title:** Molecular mimicry in multiple sclerosis

12 **Abbreviations:** BLAST - Basic Local Alignment Search Tool; CNS – Central Nervous System; EAE-

13 Experimental Autoimmune Encephalomyelitis; EBV – Epstein Barr Virus; HERV – Human Endogenous

14 Retrovirus; IEDB – Immune Epitope Data Base; MBP – Myelin Basic Protein; MOG - Myelin

15 Oligodendrocyte Glycoprotein; MS – Multiple Sclerosis; MSR - Multiple Sclerosis Associated

16 Retrovirus; NCBI – National Centre for Biological Information; PLP - Proteolipid Protein; SMM -

17 Stabilised Matrix Method.

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23 **Abstract**

24 Multiple sclerosis is an autoimmune disease caused by the destruction of the myelin  
25 sheath in the central nervous system by T cells and antibodies. The major target molecules are the  
26 myelin basic protein, the myelin oligodendrocyte glycoprotein and the proteolipid protein but the  
27 aetiology of the disease is as yet poorly understood. The HLA Class II allele DRB1\*1501 in particular  
28 as well as DRB5\*0101 and the expression of human endogenous retroviral envelope proteins have  
29 been linked to multiple sclerosis but the molecular mechanisms relating these remain to be  
30 elucidated. We hypothesised that cross-reactive peptide epitopes in the retroviral envelope proteins  
31 and myelin proteins that can be presented by the two Class II DR molecules may play a role in  
32 initiating multiple sclerosis. As an initial step to test the hypothesis, sequence homologies between  
33 retroviral envelope and myelin proteins and *in silico* predictions of peptides derived from them that  
34 are able to bind to the two Class II alleles were examined. The results support the hypothesis that  
35 molecular mimicry in peptide epitopes from envelope proteins of the HERV-W family of endogenous  
36 retroviruses and myelin proteins is possible and this may potentially trigger multiple sclerosis.  
37 Mimicry between syncytin-1, a HERV-W envelope protein that is expressed during placentation, and  
38 myelin proteins may also explain the higher prevalence of multiple sclerosis in women. Experimental  
39 confirmation of the ability of the identified peptide epitopes to activate T<sub>H</sub> cells is a logical extension  
40 to the present findings, and can lead to new immunotherapeutic procedures to treat multiple  
41 sclerosis.

42 **Key Words:** autoimmunity, human endogenous retroviruses, molecular mimicry, multiple sclerosis,  
43 myelin proteins.

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## 47 1. Introduction

48 Multiple sclerosis (MS) is an inflammatory autoimmune disease of the central nervous  
49 system (CNS) that involves the progressive destruction of the myelin sheath and axons resulting in  
50 neurodegeneration [1]. Antibodies and T cells are both considered to be responsible effectors [2, 3]  
51 and this is consistent with sequences of T and B cell antigen receptors in the CNS of MS patients [2].  
52 Experimental autoimmune encephalomyelitis (EAE) in rodents mimics many aspects of MS and is  
53 considered to be a useful animal model of MS. Studies on EAE have indicated the main target  
54 molecules in myelin for the autoimmune responses in MS. EAE can be induced in naïve mice by  
55 active immunization with the myelin basic protein (MBP), proteolipid protein (PLP) or myelin  
56 oligodendrocyte glycoprotein (MOG), and peptides derived from them, or by passive transfer of  
57 immune T cells from animals with EAE [4].

58 Large, multi-population Genome-Wide Association Studies showed that amongst all  
59 MHC alleles, the MHC Class II allele DRB1\*15:01 had the strongest association with MS [5].  
60 DRB1\*15:01 is also associated with early onset of MS [6]. DRB1\*1501 is one of the alleles coding for  
61 the DR  $\beta$  chain. The DRB1\*1501  $\beta$  chain pairs with the relatively non-polymorphic HLA DR  $\alpha$  chain  
62 from DRA\*0101 to form the HLA DR2b heterodimer. Furthermore, DRB5\*0101, which is in linkage  
63 disequilibrium with DRB1\*1501, is also associated with MS [7, 8] and binds to DRA\*0101 to form the  
64 HLA DR2a heterodimer. Both HLA DR2a and DR2b, often referred to as representing the HLA DR2(15)  
65 haplotype, have been shown present the encephalitogenic MBP peptide 83-99 to T cell clones from  
66 MS patients [7, 8]. HLA DR2a and DR2b may also present other peptide epitopes associated with MS  
67 to CD4+ T cells. Evidence suggests that CD4+ T cells with a T<sub>H</sub>1 phenotype have a central role in the  
68 initiation of MS and its progression, the generation of autoantibodies and self-reactive CD8+ T cells,  
69 and MS-associated inflammation [9, 10]. HLA DR2a and DR2b restricted T<sub>H</sub>1 cells are also able to  
70 directly lyse target cells through the perforin or Fas/FasL pathway [11]. Proinflammatory CD4+ T<sub>H</sub>17  
71 are also important in the pathogenesis of MS [12, 13].

72 Besides genetic factors, viral infections have been associated with MS and three  
73 mechanisms that are not mutually exclusive have been proposed to explain this [3]. Firstly, viral  
74 infection of the CNS can potentially cause inflammation and damage oligodendrocytes that produce  
75 myelin, thereby releasing myelin fragments that activate autoreactive T cells in an inflammatory  
76 milieu. Subsequent epitope spreading can produce more demyelination and axon death. Secondly,  
77 persistent viral infection of the CNS can produce inflammatory demyelinating disease caused by the  
78 immune response attempting to eliminate infected cells within the CNS. Thirdly, viral infection  
79 outside the CNS can activate cross-reactive T cells that then enter the CNS and cause inflammatory  
80 demyelinating lesions leading to MS. Accumulating evidence suggests that human endogenous  
81 retroviruses (HERVs), generally inactive remnants of exogenous retroviruses that became integrated  
82 into primate genomes, are important in the aetiology and pathogenesis of MS [14]. An MS  
83 associated retrovirus (MSRV), a member of the HERV-W family, has been particularly implicated in  
84 MS because virus particles and reverse transcriptase activity are detected in MS patients [14 - 16]. A  
85 role in MS pathogenesis has been ascribed to the MSRV *env* gene product which is nearly identical to  
86 syncytin-1 of the HERV-W family [15]. HERV-W encoded syncytin-1, itself a viral envelope protein  
87 remnant and a membrane glycoprotein, has evolved to perform an essential fusion function in  
88 forming the placental syncytiotrophoblast layer in humans [17, 18]. Syncytin-1 is highly conserved  
89 between members of the HERV-W family including MSRV [19]. Syncytin-1 is homologous to syncytin-  
90 2, another fusogenic envelope glycoprotein encoded in a different HERV family *viz.* HERV FRD, which  
91 has also evolved to play an important role in forming the syncytiotrophoblast [18]. The expression  
92 of the MSRV *env* gene product is significantly higher in brain lesions in MS plaques and correlates  
93 with the extent of active demyelination and inflammation [15, 16]. Furthermore, the observed  
94 temporal relationship between Epstein Barr Virus (EBV) infection and MS has been ascribed to  
95 activation of the expression of MSRV envelope protein/HERV-W syncytin-1 by EBV infection [16].

96 We hypothesised, based on the existing data, that the presentation of peptide epitopes  
97 derived from syncytin-1 or the MSRV envelope protein that cross-react with epitopes from myelin

98 proteins to CD4+ T cells in the context of the MHC Class II DR2b and DR2a molecules may be a  
99 molecular mimicry trigger for MS. We therefore determined *in silico* the potential for cross-reactive  
100 epitopes between syncytin-1, syncytin-2 and the MSRV envelope protein on one hand and human  
101 MBP, MOG and PLP on the other, that can be presented to CD4+ T cells by HLA DR2a and DR2b  
102 molecules on antigen presenting cells.

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## 104 **2. Materials and Methods**

### 105 **2.1 Sequence homologies between HERV-W and HERV FRD envelope proteins and the myelin** 106 **proteins MBP, MOG and PLP**

107 The predicted protein coding sequences of the 538 amino acid (aa) HERV-W syncytin-1  
108 (Uniprot accession number Q9UQF0), the 538 aa HERV FRD syncytin-2 (NP\_997465), the 542 aa  
109 MSRV envelope protein (AAK18189.1), the 304 aa human MBP (P02686.3), the 247 aa human MOG  
110 (Q16653.2), and the 277 aa human PLP (P60201.2) were obtained from the NCBI data base. Amino  
111 acid sequences of the proteins were compared by pairwise Basic Local Alignment Search Tool  
112 (BLAST) analysis performed online using default parameters (<https://www.ncbi.nlm.nih.gov/blast>).

### 113 **2.2 Prediction of peptides in HERV envelope proteins and myelin proteins potentially binding to** 114 **HLA DR2a and HLA DR2b molecules**

115 Syncytin-1, Syncytin-2 and the MSRV envelope protein and the three myelin proteins  
116 were analysed for peptides potentially binding to HLA DR2a and HLA DR2b molecules using the  
117 Immune Epitope Data Base or IEDB ([www.iedb.org](http://www.iedb.org)) procedures [20 - 23]. The default peptide length  
118 of 15 aa was used in the analysis but the results also show the core nonamer peptides that are  
119 expected to bind to the HLA DR molecule and constitute the major portion of the T cell epitope [20].  
120 The analysis method selected was the Stabilised Matrix Method (SMM) where the peptides are  
121 ranked according to their predicted binding affinities or IC<sub>50</sub> which indicates the concentration of

122 peptide in nM expected to achieve 50% saturation of the HLA molecule. Therefore a lower  $IC_{50}$   
123 indicates higher affinity. As a guide, peptides with  $IC_{50}$  values <50 nM are considered to bind with  
124 high affinity, between 50nM to 500 nM with intermediate affinity and between 500nM to 5000 nM  
125 with low affinity [21 - 23]. For each peptide, a percentile rank is generated by comparing the  
126 peptide's score against the scores of five million random 15 mers selected from the SWISSPROT  
127 protein database. Therefore smaller percentile rank values, typically <10, also indicate higher affinity  
128 and specificity of binding to the HLA molecule [21 - 23].

129           The predicted peptides binding with higher-affinity in myelin proteins were then  
130 examined to determine whether they were located in regions that were homologous to syncytin-1,  
131 syncytin-2 and the MSRV envelope protein. Homologies between the predicted syncytin-1, syncytin-  
132 2 and the MSRV envelope protein and myelin protein peptides were additionally tested by pairwise  
133 BLAST analysis of the peptides.

### 134 **3. Results**

#### 135 **3.1 Sequence homologies between HERV envelope proteins and the three myelin proteins**

136           A comparison of the syncytin-1 and MSRV envelope protein by BLAST revealed that the  
137 two proteins were 87% identical with 90% positives and 4 gaps (Supplementary Figure S1). A  
138 comparison of syncytin-1 with syncytin-2 showed that these two proteins were more distantly  
139 related with significant homology present only between parts of the two proteins (Figure S1b). The  
140 results obtained with pair-wise BLAST analysis of syncytin-1 and MBP, MOG, and PLP are presented  
141 in Supplementary Figures S2, S3 and S4 respectively. The results obtained with pair-wise BLAST  
142 analysis of MSRV envelope protein with MBP, MOG, and PLP are presented in Supplementary  
143 Figures S5, S6 and S7 respectively

144           There are three regions of homology between syncytin-1 and MBP, with greatest  
145 homology of 29% with an E value of 0.07 being found between aa 223-274 of MBP and aa 8-58 of

146 syncytin-1 allowing for a total of nine gaps in both proteins (Figure S2). The region between aa 223-  
147 274 of MBP was also homologous to MSRV envelope protein aa 8-58, with an E value of 1.8 and nine  
148 gaps (Figure S5). Two other regions of homology identified between syncytin-1 and MBP possessed  
149 weaker homologies with E values of 0.7 and 6 (Figure S2). Similarly two other regions of weaker  
150 homology were detected between MBP and MSRV envelope protein with E values of 2.7 and 9.4,  
151 with only the latter region being identical to a region of homology detected between MBP and  
152 syncytin-1 (Figure S5).

153           A short region of significant homology of 62% identity without gaps with an E value of  
154 0.03 was seen between MOG aa 214-226 and syncytin-1 aa 448-460 (Figure S3). However this region  
155 was not detected in the pairwise BLAST comparison of MOG and MSRV envelope protein but four  
156 other regions of weak homologies with E values >2.3 were detected (Figure S6).

157           Between syncytin-1 and PLP, only one region of weak homology was identified viz.  
158 between aa 114-154 of PLP and aa 411-453 of syncytin-1 with 28% identity and an E value of 5.9  
159 allowing for two gaps in the PLP sequence (Figure S4). The corresponding region of MSRV envelope  
160 protein aa 411-464 was also homologous to PLP aa 114-167 with 29% identity, E value of 0.9 and  
161 four gaps (Figure S7).

162           No regions of significant homology were observed between syncytin-2 and the three  
163 myelin proteins (data not shown).

### 164 **3.2 Peptides in HERV envelope proteins and the three myelin proteins predicted to bind to HLA** 165 **DR2b molecules**

166           The results of IEDB analysis of syncytin-1, MSRV envelope protein, MBP, MOG and PLP  
167 binding HLA DR2b are shown in Supplementary Tables S8, S9, S10, S11 and S12 respectively.

#### 168 **3.2.1 Syncytin-1**

169                    There were twelve 15mer peptides in syncytin-1 with an  $IC_{50} < 50$  nM and percentile  
170 rank  $\leq 0.4$  predicted to be able to bind to the HLA DR2b molecule (Table S8). Four of these twelve  
171 high affinity predictions contained the core nonamer sequence ILPFLGPLA corresponding to aa 448  
172 to 456 in the syncytin-1 sequence that is homologous to the sequence TLPFLGPLA (aa 448-456) in  
173 the MSRV envelope protein (Figure S1). This sequence in syncytin-1 is within a short sequence  
174 significantly homologous to MOG that was identified by BLAST analysis (Figure S3). In the  
175 corresponding homologous region of MOG are two overlapping peptides with core nonamer  
176 sequences of IVPVLGPLV (aa 214-222) and ITLFVIVPV (aa 209-217) predicted to have a high affinity  
177 of binding to HLA DR2b (Table S11 and Section 3.2.4 below). The syncytin-1 nonamer ILPFLGPLA  
178 shows 6/9 positional identities with the MOG nonamer IVPVLGPLV.

### 179 **3.2.2 MSRV envelope protein**

180                    The MSRV envelope protein was predicted to have 10 peptides with high affinity to HLA  
181 DR2b (Table S9). Four, with nonamer sequence of FRPYISIPV (aa 194-202) and four with the nonamer  
182 sequence LVKFVSSRI (aa 472-480) were homologous to the predicted high affinity sequences  
183 FRPYVSIPV (aa 194-202) and LVNFYSSRI (aa 472-480) respectively in syncytin-1 (the different amino  
184 acids in the nonamers are underlined). Two other predicted nonamers of high affinity to the MSRV  
185 envelope protein *viz.* LPLHFRPYI (aa 190-198) and FNELVKFVS (aa 469-477) show two differences  
186 from the syncytin-1 high affinity nonamers LPLNFRPYV (aa 190-198) and FNLLVNFVS (aa 469-477)  
187 respectively.

188                    The nonamer with sequence TLPFLGPLA (aa 448-456) in the MSRV envelope protein  
189 that is homologous to high affinity nonamer sequences in syncytin-1 and MOG (section 3.2.1) lies  
190 within a MSRV envelope protein peptide predicted to have an intermediate affinity to HLA DR2b  
191 (Table S9).

### 192 **3.2.3 MBP**

193 IEDB analysis of MBP showed that there were five 15mer peptides that were predicted  
194 to bind with high affinity to HLA DR2b and with a percentile rank  $\leq 0.14$  (Table S10). The core  
195 nonamer peptides for these were all derived from residues 220/221 to 228/229 with the sequences  
196 (V)VHFFKNIV(T) that did not show detectable homology with syncytin-1, syncytin-2 or the MSRV  
197 envelope protein.

### 198 **3.2.4 MOG**

199 IEDB analysis of MOG predicted five peptides with high affinity of for HLA DR2b and percentile rank  
200  $\leq 0.34$  (Table S11). The homology of the MOG overlapping core nonamer sequence IVPVLGPLV  
201 (residues 214-222) from these with the predicted syncytin-1 core nonamer ILPFLGPLA and the MSRV  
202 nonamer TLPFLGPLA has been described in section 3.2.1. Syncytin-2 did not possess a homologous  
203 nonamer to the MOG nonamer IVPVLGPLV.

### 204 **3.2.5 PLP**

205 IEDB analysis of PLP showed that there were five 15mer peptides predicted to have a  
206 high affinity of binding to HLA DR2b with percentile rank  $\leq 0.43$  (Table S12). These had core nonamer  
207 sequences of which one FFFLYGALL (aa 78-86) showed homology with a syncytin-1/MSRV envelope  
208 protein sequence FLFTVLL (aa 8-14) with 4/7 identities and an E value of 0.19 (Figure S12b). The  
209 sequence FLFTVLL appeared in many syncytin-1 and MSRV envelope protein peptides that were  
210 predicted to have an intermediate affinity for HLA DR2b (Tables S8 and S9) but syncytin-2 did not  
211 possess a homologous sequence.

### 212 **3.2.6 Syncytin-2**

213 The four nonamers in syncytin-2 that were predicted to have a high affinity of binding  
214 to HLA DR2b were located within the signal peptide and not homologous to the three myelin  
215 proteins (data not shown).

### 216 **3.3 Peptides in HERV envelope proteins and the three myelin proteins predicted to bind to HLA**

#### 217 **DR2a molecules**

218                   The results of IEDB analysis of syncytin-1, MSRV envelope protein, MBP, MOG and PLP  
219 peptides binding to HLA DR2a are shown in Supplementary Tables S13, S14, S15, S16 and S17  
220 respectively.

#### 221 **3.3.1 Syncytin-1**

222                   There were 14 peptides (15mers) in syncytin-1 with a predicted high affinity of binding  
223 to HLA DR2a. Nine of these peptides had an affinity of  $<10\text{nM}$   $\text{IC}_{50}$  with a percentile rank = 0.01.  
224 However none of the core nonamers derived from these sequences showed a significant homology  
225 with the three myelin proteins.

#### 226 **3.3.2 MSRV envelope protein**

227                   One peptide with a predicted high affinity to HLA DR2a containing the core nonamer  
228 FLVKFVSSR (aa 471-479) was identified in the MSRV envelope protein. However this was not  
229 significantly homologous to sequences within the three myelin proteins in BLAST analysis.

#### 230 **3.3.3 MBP, MOG and PLP**

231                   Seven 15mer peptides from MBP, five 15mers from MOG and seventeen 15mers from  
232 PLP were predicted to bind to HLA DR2a with high affinity and percentile ranks  $\leq 0.13$ , 0.22 and 0.23  
233 respectively. However none of the core nonamers derived from these peptides were significantly  
234 homologous to sequences within syncytin-1, syncytin-2 or the MSRV envelope protein.

#### 235 **3.3.4 Syncytin-2**

236                   In syncytin-2, the ten nonamers with predicted high affinity for HLA DR2a did not show  
237 significant homology to the three myelin proteins (data not shown).

### 238 **3.4 MBP peptide that binds to both HLA DR2a and HLA DR2b**

239           The encephalitogenic MBP peptide 83-99 that had been shown to be presented in the  
240 context of HLA DR2a and HLA DR2b to T cell clones from MS patients [7, 8], was shown in the IEDB  
241 analysis to lie in a region predicted to have an intermediate affinity of binding to both HLA DR2b and  
242 DR2a and with percentile ranks of  $\leq 2.36$  and  $\leq 2.84$  respectively (Tables S10 and S15), consistent with  
243 the validity of the IEDB analytical procedure used here. However MBP 83-99 is in a region that was  
244 not homologous to syncytin-1 and the MSRV envelope protein by the BLAST analysis (Figures S2 and  
245 S5 respectively) or to syncytin-2 (data not shown).

### 246 **3.5 Immunodominant peptides in MOG recognised in the context of HLA DR2(15)**

247           Among synthetic peptides specifically able to stimulate T cells from MS patients, the  
248 MOG aa 63-87 peptide was shown to be immunodominant in an independent study [24]. The IEDB  
249 analysis showed that this peptide region has a high affinity for HLA DR2a and an intermediate affinity  
250 for HLA DR2b (Tables S11 and S16) consistent with the experimental findings. However MOG 63-87  
251 was not detectably homologous to syncytin-1 and the MSRV envelope protein in BLAST analysis  
252 (Figures S3 and S6 respectively) or to syncytin-2 (data not shown).

253           Another human MOG peptide aa 35-55 was shown to be immunogenic in the context of  
254 HLA DR2b in transgenic mice and therefore potentially encephalitogenic in humans [25]. This region  
255 of MOG is predicted to be have weak affinity for HLA DR2b [ $IC_{50}$  524 nM, percentile rank 10.21, core  
256 nonamer 32-40 FRVIGPRHP in Table S11] and intermediate affinity for HLA DR2a [ $IC_{50}$  57 nM,  
257 percentile rank 0.3, with the same core nonamer 32-40 FRVIGPRHP in Table S16] in the IEDB  
258 analysis. However this region of MOG was not detected as being homologous to syncytin-1 and the  
259 MSRV envelope protein by BLAST analysis [Figures S3 and S6 respectively] or to syncytin-2 (data not  
260 shown).

### 261 **3.6 PLP peptides recognised in the context of HLA DR2(15)**

262                   The PLP peptide 175-194 has been shown to be presented by transgenic murine T cells  
263 in the context of HLA DR2b [26] and this is consistent with the intermediate level of affinity for HLA  
264 DR2b and HLA DR2a observed in our present analysis (Tables S12 and S17 respectively). However  
265 this region of PLP was not significantly homologous to sequences in the two HERV-W envelope  
266 proteins (Figures S4 and S7) or to syncytin-2 (data not shown).

267

## 268 **Discussion**

269                   The IEDB analyses identify several peptides in the two HERV-W envelope proteins that  
270 can be potentially presented to T<sub>H</sub> cells with high affinity (<50nM IC<sub>50</sub>) by HLA DR2b (12 peptides)  
271 and HLA DR2a (14 peptides), two Class II molecules that have been significantly associated with MS  
272 [5 – 8]. Some syncytin-1 peptides predicted to bind to HLA DR2a were expected to do so with  
273 particularly high affinity. Many potential binding peptides in the envelope proteins with  
274 intermediate (50-500nM IC<sub>50</sub>) and weak (500-5000nM IC<sub>50</sub>) affinities for HLA DR2b and HLA DR2a  
275 were also identified in the analyses. The findings are therefore consistent with the hypothesis that  
276 HERV-W envelope proteins contain many potential epitopes for CD4+ T cells that can be presented  
277 by MS- associated Class II HLA-DR molecules on antigen presenting cells.

278                   The IEDB analyses also identified several peptides in the three myelin proteins that are  
279 predicted to bind with high affinity to HLA DR2b and HLA DR2a. As in the case of the HERV-W  
280 envelope proteins, many other potential peptides with intermediate (50n-500nM IC<sub>50</sub>) and weak  
281 (500-5000nM IC<sub>50</sub>) affinities for HLA DR2b and HLA DR2a were also identified in the three myelin  
282 proteins. The results show the potential for many peptides from the three myelin proteins to be  
283 presented to T<sub>H</sub> cells in the context of HLA DR molecules known to be associated with a  
284 predisposition to MS. Furthermore, peptides from the three myelin proteins that had been shown  
285 experimentally to be recognised by T<sub>H</sub> cells in the context of presentation by HLA DR2b and HLA

286 DR2a molecules, were shown by the *in silico* analysis here to have the potential for presentation by  
287 HLA DR2b and HLA DR2a.

288           The identification of peptides from syncytin-1 and the MSRV envelope protein that bind  
289 with a predicted high or intermediate affinity respectively to HLA DR2b that also show a degree of  
290 sequence homology to predicted high affinity peptides derived from MOG is consistent with the  
291 molecular mimicry hypothesis for MS. The identification of a PLP peptide with a predicted high  
292 affinity for HLA DR2b that shows sequence homology with predicted peptides from the two HERV-W  
293 envelope proteins with intermediate affinity for HLA DR2b is also in keeping with this hypothesis.  
294 The observation that immunisation with the MSRV envelope protein can induce EAE in mice is  
295 supportive of the hypothesis [27].

296           The question arises as to how syncytin-1 which plays a normal role in foetal  
297 development and also reported to be involved in maintaining maternal tolerance to the foetus [17-  
298 19] may become immunogenic in the context of MS. Syncytin-1 and Isyncytin-2 are membrane  
299 glycoprotein found in the plasma membrane of trophoblast cells that are also shed as exosomes into  
300 the maternal circulation [19]. The MSRV envelope glycoprotein, which is nearly identical to syncytin-  
301 1, is on the outer membrane of complete viral particles [18] that have been detected in MS lesions in  
302 the CNS [14-16], including in monocytes and macrophages [16]. It is possible therefore syncytin-1 in  
303 circulating exosomes and MSRV envelope protein whose expression may be induced within or  
304 outside the CNS, can be taken up by antigen presenting cells, processed and presented to T<sub>H</sub> cells on  
305 HLA DR2b or HLA DR2a molecules. Antigen presenting cells in the CNS include microglia, astrocytes  
306 and perivascular monocytes and macrophages and also inflammation-activated endothelium [28].  
307 MSRV envelope protein has been shown to activate human monocytes via the pattern recognition  
308 receptors CD14 and TLR4 to induce the formation of the proinflammatory cytokines TNF $\alpha$ , IL-1 $\beta$  and  
309 IL-6, and to activate dendritic cells to promote T<sub>H</sub>1 responses [29]. It is therefore possible that HERV-  
310 W envelope proteins in the form of syncytin-1 or the MSRV envelope protein are a source of peptide

311 epitopes that can initially activate T<sub>H</sub> cells in the context of HLA DR2b or HLA DR2a, particularly in an  
312 inflammatory context. One source of inflammation can be the activation of MSR<sub>V</sub> transcription to  
313 produce MSR<sub>V</sub> envelope protein by infections such as EBV [16]. A proportion of the activated T<sub>H</sub> cells  
314 may then be expected to react with the cross-reactive MOG and PLP peptide epitopes predicted  
315 here that are processed and presented by CNS resident antigen presenting cells on HLA DR2b  
316 molecules. The resulting amplified inflammation, recruitment of cytotoxic T cells and formation of  
317 anti-MOG and anti-PLP antibodies can further damage oligodendrocytes that make myelin and the  
318 myelin sheath on nerve cells. This can result in release of more myelin proteins that can be taken up  
319 by antigen presenting cells to cause epitope spreading and activation of additional T<sub>H</sub> cells.  
320 Additional myelin protein – derived peptides may be presented in the context of HLA DR2b or HLA  
321 DR2a as predicted here or in the context of other HLA molecules as has been experimentally shown  
322 to be possible [30] to result in full-blown MS. We therefore suggest that the *in silico* analysis  
323 provides support for molecular mimicry between peptides derived from HERV-W envelope proteins  
324 and myelin proteins presented by HLA DR2b and HLA DR2a helping to initiate MS. However the  
325 available data does not support a similar role for HERV FRD encoded syncytin-2 in MS. Experimental  
326 investigations on the ability of the peptides identified in this study to be recognised by antigen  
327 receptors on T<sub>H</sub> cells in the context of HLA DR2b and HLA DR2a, and then become appropriately  
328 activated, are needed to confirm the hypothesis.

329           The identification of important peptide epitopes can lead to the development of small  
330 molecule inhibitors that impede their presentation by HLA DR2b and HLA DR2a to treat MS in its  
331 early stages [31]. Two other relevant immunological approaches to treat MS that have shown  
332 promise in the murine EAE model include infusing syngeneic antigen presenting cells containing  
333 chemically coupled immunogenic MOG peptide which leads to induction of regulatory T<sub>H</sub> cells,  
334 downregulation of inflammatory T<sub>H</sub> cells and reversal of EAE [32, 33], as well as immunisation with  
335 cryptic peptide epitopes from MOG that ameliorates EAE [34].

336 Women have a higher prevalence of MS than men during and after the child bearing  
337 age and this gender difference has been attributed to hormonal effects and differential  
338 environmental exposure to as yet undetermined agents [35, 36]. It has been reported that MSR  
339 copy numbers are increased in female MS patients compared to male patients, that this increase  
340 correlates with disease severity [36], and that the increase may be caused by HERV-W copies in the X  
341 chromosome [37, 38]. Our findings suggest that the expression of syncytin-1 during pregnancy and  
342 molecular mimicry between syncytin-1 and myelin proteins may contribute, at least in part, to the  
343 higher prevalence of MS among women.

#### 344 **Conflict of interest statement**

345 The authors declare no conflict of interest.

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#### 349 **Author contributions**

350 TW and RR designed the project, BJ performed the initial analysis, and RR carried out  
351 detailed analysis and drafted the manuscript. All authors read and approved the final manuscript.

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