1 Coastal reservoirs as a source of nitrous oxide: Spatio-temporal

2 patterns and assessment strategy

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$25 \quad \mathbf{ABSTRACT}$

Coastal reservoirs are widely regarded as a viable solution to the water scarcity 26 27 problem faced by coastal cities with growing populations. As a result of the accumulation of anthropogenic wastes and the alteration of hydroecological processes, 28 29 these reservoirs may also become the emission hotspots of nitrous oxide (N2O). Hitherto, accurate global assessment of N2O emission suffers from the scarcity and 30 low spatio-temporal resolution of field data, especially from small coastal reservoirs 31 with high spatial heterogeneity and multiple water sources. In this study, we measured 32 33 the surface water N2O concentrations and emissions at a high spatial resolution across three seasons in a subtropical coastal reservoir in southeastern China, which was 34 hydrochemically highly heterogeneous because of the combined influence of river 35 36 runoff, aquacultural discharge, industrial discharge and municipal sewage. Both N2O concentration and emission exhibited strong spatio-temporal variations, which were 37 correlated with nitrogen loading from the river and wastewater discharge. The mean 38 39 N2O concentration and emission were found to be significantly higher in the summer than in spring and autumn. The results of redundancy analysis showed that NH4+-N 40 41 explained the greatest variance in N2O emission, which implied that nitrification was the main microbial pathway for N2O production in spite of the potentially increasing 42 importance of denitrification of NO3--N in the summer. The mean N2O emission 43 across the whole reservoir was 107 µg m-2 h-1, which was more than an order of 44 magnitude higher than that from global lakes and reservoirs. Based on our results of 45 Monte Carlo simulations, a minimum of 15 sampling points per km2 would be needed 46

to produce representative and reliable N2O estimates in such a spatially
heterogeneous aquatic system. Overall, coastal reservoirs could play an increasingly
important role in future climate change via their N2O emission to the atmosphere as
water demand and anthropogenic pressure continue to rise. *Keywords:* Nitrous oxide (N₂O); Spatial heterogeneity; Spatially resolved

52 measurement; Wastewater discharge; Subtropical reservoir; IPCC

53 **1. Introduction**

Nitrous oxide (N₂O) is a potent greenhouse gas with a global warming potential 54 55 nearly 300 times that of carbon dioxide (CO₂) on a mass basis over a 100-year time horizon (IPCC, 2013). N₂O is also considered to be one of the major substances that 56 57 can destroy ozone in the stratosphere (Ravishankara et al. 2009; Shaaban et al., 2018). The atmospheric averaged N₂O concentration reached 331.1 ± 0.1 ppbv in 2018, which 58 is approximately 23% higher than the pre-industrial (before 1750) levels (World 59 Meteorological Organization, 2019). There has been a steady increase in atmospheric 60 N_2O at a rate of 0.7–0.8 ppb yr⁻¹ over the past three decades (Davidson 2009; 61 Saikawa et al. 2014; Xiao, et al., 2019a). Quantifying the potential source strength of 62 various ecosystems is fundamental for predicting future N₂O emission and climate 63 64 change (Yang et al., 2020).

Aquatic ecosystems are considered as important sources of N₂O emission, 65 contributing for approximately 25%–30% of global N₂O emissions (Zhou et al., 2019). 66 67 As an important component of the Earth's surface water systems, man-made reservoirs, which include those for hydropower, flood management, water supply, and navigation 68 purposes, are now considered as significant contributors of the atmospheric N₂O 69 (Beaulieu et al., 2019; Cheng et al., 2019; Guérin et al., 2008; Wang et al., 2017; Yang 70 & Flower 2012). Recent estimates indicated that the amount of N₂O emitted from 71 global reservoirs was approximately 0.03-0.07 Tg N yr⁻¹ (Deemer et al., 2016; 72 Maavara et al., 2019). However, the estimate of N₂O emission from reservoirs, 73 especially from the ones with anthropogenic disturbances, remains highly uncertain 74

due to the data limitation. Additionally, N₂O emissions from reservoirs in different 75 climatic zones exhibited high spatial-temporal variations across and within the 76 77 systems (Beaulieu et al., 2015; Cheng et al., 2019; Liang et al., 2019; Liu et al., 2011a, 2015, 2017; Musenze et al., 2014; Shi et al., 2020). This is another source of 78 uncertainty in reservoir N₂O budget. Thus, more in situ measurements with high 79 spatiotemporal resolution are needed to reduce the uncertainty and to develop more 80 accurate approaches for upscaling to whole reservoir N₂O emissions and further 81 large-scale assessments of reservoir N₂O fluxes. 82

83 The N₂O production in reservoir is known to mainly derive from the nitrification and denitrification of terrestrial nitrogen (N). Human activities in the drainage basin, 84 particularly land use change, sewage discharge and agriculture fertilization, have 85 86 severely altered the N transport of from the terrestrial ecosystem to the aquatic ecosystem (Davidson, 2009; Hosen et al., 2014; Williams et al., 2016), and influenced 87 the N₂O production in reservoirs (Wang et al., 2017). Syvitski et al. (2005) estimated 88 89 over 100 billion metric tons of sediment and up to 3 billion metric tons of organic matter were trapped in the reservoirs in the last five decades. Hence the allochthonous 90 91 material load by human activities would play a much more important role for accelerating N₂O emission from reservoirs. To date, some efforts have been made on 92 the response of N₂O emission from reservoirs to the human activities (e.g., agriculture, 93 urbanization sewage discharges) in the basin, especially in China (e.g., Liu et al., 94 2011a; Wang et al., 2017; Xiao et al., 2019b). However, the data of N₂O emissions 95 from reservoirs under the impact of human activities are disproportionately scarce in 96

comparison to the amount and area of reservoirs. More importantly, the majority of existing studies only focused on inland freshwater reservoirs, but few studies have mentioned N₂O emission from coastal reservoirs. As coastal reservoirs often experience a higher salinity and inputs of anthropogenic carbon and nutrients than inland ones, N₂O production and emission rates might differ between the reservoirs in these two zones. However, there is currently a lack of empirical data to test this hypothesis and advance our understanding of reservoirs in the global N₂O cycle.

In order to fill these knowledge gaps, this study researched high spatial 104 105 resolution measurements of dissolved N₂O concentrations in a subtropical coastal reservoir in southeast China over three seasons of a year to estimate the N₂O fluxes 106 across the water-atmosphere interface. The specific objectives of this study are to (1) 107 108 quantify the magnitude of N₂O emission fluxes from the coastal reservoirs, (2) assess the spatial variations in diffusive N₂O fluxes both within system and among systems, 109 (3) explore the drivers of the spatial variations of coastal reservoir N_2O emissions; 110 111 and (4) examine how many measurements sities in space were needed to representatively cover the observed spatial variability in coastal reservoir N2O 112 emissions. The results of the present study will provide the scientific basis for the 113 development of global biogeochemical models and national GHG inventories through 114 characterizing N₂O emission from the reservoirs in coastal areas. 115

- 116 **2. Materials and Methods**
- 117 *2.1. Study Area*

118

The study was conducted in Wenwusha Reservoir (25°49'36"-25°54'00" N,

119 $119^{\circ}35'12''-119^{\circ}38'11''$ E), a subtropical reservoir in the coastal area of Fujian 120 Province, Southeast China (Figure 1). The reservoir was built for irrigation and 121 flood-prevention. It has a surface area of 5.2 km², a total volume of 3.20×10^8 m³, 122 drainage area of 275 km², and mean depth of 9.0 m. The reservoir is influenced by a 123 subtropical monsoon humid climate, with a mean annual temperature of 19.3 °C. The 124 average annual precipitation is 1390 mm, most of which (approximately 75%) occurs 125 in the wet season from May to September.

The Wenwusha Reservoir was created by two dams on the Nangyangdong River 126 estuary. We compartmentalized the reservoir into two reservoir zones according to the 127 two dam constructed time, topographic feature, and trophic status (Figure 1). The 128 northern reservoir zone (NRZ) was constructed in 1957, with a surface area of 1.9 129 km^2 and a total volume of $1.40 \times 10^8 m^3$. The NRZ is located in highly urbanized 130 districts and is heavily impacted by human activities, e.g. the sewage discharge from 131 aquacultural, industrial, and municipal activities. The NRZ also received large 132 amounts of nutrient input from Nanyangdong river. The southern reservoir zone (SRZ) 133 was constructed in 2004, with a surface area of 3.3 km² and a total volume of 134 1.69×10^8 m³. Around 70% of its catchment is used for agricultural activities (e.g., 135 aquaculture and farming), but its immediate surroundings are partly forested and 136 wetland (Figure 1). The water salinity in NRZ is much lower than that in SRZ 137 (0.4–1.3‰ versus 0.4–3.7‰), as a result of freshwater dilution caused by the surface 138 runoff. 139

140 2.2. Sampling strategies

141	Considering the reservoir size, topographic feature, and pollution source, a total
142	of 21 transects (10 in the NRZ and 11 in the SRZ) were selected to collect samples to
143	characterize the variation across the whole reservoir. According to the lengths of the
144	transects, there were 3-10 sampling sites in the transects (Figure 1). The total number
145	of sample sites was 103 (56 in the NRZ and 47 in the SRZ). These sites were
146	distributed in 5 different sewage loading sectors, namely the industrial effluent
147	loading sector (Sector-I, $n = 4$), town sewage loading sector (Sector-T, $n = 6$), river
148	input sector (Section-R, $n = 7$), aquaculture sewage loading sector (Section-A, $n = 22$)
149	and non-wastewater loading sector (Sector-N, $n = 64$). At each site, surface-water
150	samples were collected at a depth of 20 cm. Three in situ whole reservoir surveys
151	were conducted in mid–November 2018, mid–March and mid–June 2019. Each whole
152	reservoir survey was completed in two consecutive days to reduce the biases caused
153	by day-to-day variation. More details about the sampling transects are presented in
154	supporting information Table S1.

155 2.3. Measurement of dissolved N₂O concentration and diffusive flux

156 2.3.1. Dissolved N₂O concentration

157 Surface water samples for dissolved N₂O concentration analysis were collected 158 using a 100-mL syringe equipped with three-way stopcocks. After sample was 159 collected from reservoir, water was quickly transferred into a 55-mL gas-tight glass 160 serum bottle. Prior to sealing the bottle, 0.2 mL saturated HgCl₂ solution was injected 161 into the sample bottle for inhibiting microbial activity. The bottle was immediately 162 sealed with an open-topped screw cap equipped with a halobutyl rubber septum to exclude any air bubble (Borges et al., 2018; Xiao et al., 2019a; Yang et al., 2020). All
water samples were stored in an ice box, transported to the laboratory within 6 hr, and
analyzed within two days of collection.

Dissolved N₂O concentrations in the collected water samples were measured 166 167 following the headspace equilibration method described by Yu et al. (2013, 2017). In the laboratory, ultrahigh purity N₂ gas (99.999%) was injected into the sample bottle 168 via a syringe to create a 25-mL headspace. The bottles were then shaken vigorously 169 for 10 min in an oscillator to allow the dissolved N₂O diffuse out and gases reach 170 equilibrium between the liquid phase and headspace. After waiting for 0.5 hr, 171 approximately 5 mL air sample was drawn from the headspace using a syringe 172 equipped with three-way stopcocks. N₂O concentrations in the headspace air were 173 174 measured using gas chromatography (GC-2014, Shimadzu, Kyoto, Japan) equipped with an electron capture detector (ECD). The detection limit for the N₂O analysis was 175 0.02 ppm, and the relative standard deviations of N₂O analyses were $\leq 5.0\%$ (Yang et 176 177 al., 2020). Based on the equilibrium temperature, salinity-dependent Henry's law constant, and the measured headspace gas concentration, dissolved N₂O concentration 178 was calculated using the formula provided by Weiss and Price (1980). 179

180 2.3.2. N₂O flux from the transfer coefficient method

The N₂O flux (F_{W-A} , µmol m⁻² h⁻¹) across the water–atmosphere interface was calculated using the classic boundary-layer model (equation (1)), which has been widely used for N₂O emissions in the lentic ecosystem (e.g., lakes, reservoir and ponds) (e.g., Cole and Caraco, 1998; Liang et al., 2019; Musenze et al., 2014; Xiao et 185 al., 2019b; Yang et al., 2015a).

186
$$F_{W-A} = k \times (C_{obs} - C_{eq})$$
(Eq1)

187 where C_{obs} is the measured dissolved N₂O concentration (nmol L⁻¹) in the surface 188 water (20 cm depth); C_{eq} in water that is in equilibrium with the atmosphere at the *in* 189 *situ* temperature (nmol L⁻¹); and *k* is the gas transfer velocity (m h⁻¹).

In the lentic ecosystem, the N₂O transfer velocity is mainly driven by wind speed because no surface water flow (Xiao et al., 2019b). k in the present study was calculated using a wind-dependent formula (equation (2)) derived from a small shallow lake as follows (Cole and Caraco, 1998):

194
$$k = (Sc/660)^{-n}(2.07 + 0.215 \times U_{10}^{1.7})$$
 (Eq2)

where n = 0.66 and 0.50 for wind speeds ≤ 3 and >3 m s⁻¹, respectively; *Sc* is the Schmidt number for N₂O and is dependent on the temperature (*T*, °C); and *U*₁₀ is the frictionless wind speed at 10 m high expressed in m s⁻¹.

198 The *Sc* and U_{10} were estimated using the following equations (Crusius and 199 Wanninkhof, 2003; Wanninkhof, 1992):

$$200 \quad Sc = 2055.6 - 137.11T + 4.3173T^2 - 0.054350T^3$$
 (Eq3)

201
$$U_{10} = U_z [1 + \frac{(C_{d10})^{1/2}}{K} \ln(\frac{10}{z})]$$
 (Eq4)

where U_z is the wind speed (m s⁻¹) at height z (2.0 m in this study) above the water surface at which wind speed was measured; C_{d10} is the drag coefficient at 10 m above the water surface (0.0013 m s⁻¹); and K is the von Karman constant (0.41).

205 2.4. Measurement of environmental variables

206 During each campaign, from 103 sites in the reservoir was also sampled for

207	measuring chemical properties. Surface water samples were collected at a depth of 20
208	cm using a 5 L organic glass hydrophore, and transferred into 150 mL polyethylene
209	bottles. Approximately 0.5 mL saturated HgCl ₂ solution was added to each bottle to
210	inhibit microbial activities (Taipale and Sonninen 2009; Zhang et al., 2013). All
211	samples were preserved in an ice box, and transported to the laboratory for
212	measurement. After filtering the samples through a 0.45 μ m cellulose acetate filter
213	(Biotrans TM nylon membranes), the concentration of total dissolved nitrogen (TDN),
214	nitrate-nitrogen (N–NO ₃ ^{$-$}), and ammonium-nitrogen (N–NH ₄ ⁺) was measured using a
215	flow injection analyzer (Skalar Analytical SAN ⁺⁺ , The Netherlands). The detection
216	limits for TDN, NN–NO ₃ ⁻ , and N–NH ₄ ⁺ were 3.0, 0.6 and 0.6 μ g L ⁻¹ , respectively.
217	The relative standard deviations of TDN, NN-NO3 ⁻ , and N-NH4 ⁺ analyses were
218	$\leq 2.0\%, \leq 3.0\%$ and $\leq 3.0\%$, respectively.

Water temperature (W_T) , dissolved oxygen (DO), pH, electrical conductivity 219 (EC), and salinity at a depth of 20 cm were also measured in situ at each sampling site. 220 $W_{\rm T}$ and pH were measured using a portable pH/mV/Temperature meter (IQ150, IQ 221 Scientific Instruments, USA). DO, EC and salinity were determined using a 222 multiparameter water quality probe (550A YSI, USA), an electrical conductivity 223 meter (2265FS EC, Spectrum Technologies, USA) and a salinity meter (Eutech 224 Instruments-Salt6, USA), respectively. The relative standard deviations of DO, pH, 225 EC, and salinity analyses were $\leq 2.0\%$, $\leq 1.0\%$, $\leq 1.0\%$ and $\leq 1.0\%$, respectively. 226 Meanwhile, the meteorological factors, including the air temperature $(A_{\rm T})$, 227 atmospheric pressure (A_P) and 2 m height wind speed (W_S) , were recorded using a 228

229 portable weather meter (Kestrel-3500, USA).

230 2.5. Statistical analysis

231 The analysis of variance (ANOVA) to test for significant (p < 0.05) effects of reservoir zone and sewage sector water depth on surface water physicochemical 232 233 properties, dissolved N₂O concentration and N₂O diffusive fluxes using the SPSS 17.0 234 statistical software package (SPSS Inc., USA). Pearson correlation coefficients were used to examine the relationships between water physicochemical parameters and 235 N₂O concentration/ fluxes. Principal component analysis (PCA) was also performed 236 to analyze the relationships between the N₂O fluxes and surface water 237 physicochemical properties to show their pattern at different sampling campaigns. For 238 quantifying spatial heterogeneity effects by randomizing the high-resolution N₂O 239 240 measurements, a Monte Carlo analysis was used to evaluate the effect of sample size on the N₂O estimation of the whole reservoir. Without replacements, the N₂O 241 measurements are resampled from the 103 sites (n = 10, 20, 30..., 100). The 242 243 resampling process then was repeated 10000 times and the overall mean and standard deviation of N₂O emissions were calculated for each sample size. The results were 244 presented as means ± SE. Statistical plots were generated using OriginPro 7.5 245 (OriginLab Corp. USA) and ArcGIS 10.2 (ESRI Inc., Redlands, CA, USA). 246 Conceptual diagrams were drawn using EDraw Max version 7.3 (EdrawSoft, Hong 247 Kong, China). 248

3. Results

250 *3.1. Water Quality Parameters in Reservoir*

251	There were substantial spatial variations in water chemical parameters (e.g.,
252	salinity, DO, and nutrient concentrations) between both reservoir zones on all three
253	sampling occasions (p <0.01; Table S2), with lower levels of DO (Figure 2a-c), EC
254	(Figure 3g-i and Figure S1d) and salinity (Figure 2j-l and Figure S1c) and higher
255	concentrations of N–NO ₃ ⁻ (Figure 2d-f and Figure S2a-c), N–NH ₄ ⁺ (Figure 2g-i and
256	and Figure S2d-f), and TDN (Figure 2j-l and Figure S2g-i) in NRZ than in SRZ.
257	However, the W_T (Figure 3a-c) and pH (Figure 3d-f) were relatively constant across
258	the reservoir ($p>0.05$; Figure S1a and b), with the coefficients of variation only ranged
259	from 4% to 7%, and 1% to 7% over the three sampling campaigns, respectively.
260	The mean DO, salinity, $N-NO_3^-$, $N-NH_4^+$ and TDN also differed significantly
261	among the five sewage loading sectors over the three sampling campaigns (Table S3).
262	Mean $N-NO_3^-$, $N-NH_4^+$ and TDN concentrations were significantly lower in
263	Sector-N than the other sectors ($p < 0.05$; Table S3), and were generally highest in
264	Sector-R. In contrast, mean DO concentration and salinity was generally greater in
265	Sector-N than the other four sectors (Table S3).
266	The main water quality parameters also showed strong temporal variation across
267	the whole reservoir (p <0.001, Two-way ANOVAs; Table S2). The water temperature
268	was highest in summer (29.3 °C), followed by spring (23.1 °C) and autumn (18.3 °C)

270 (Figure 3d-f) and salinity (Figure 3j-l), concentrations were contrary to that of water

269

(Figure 3a-c). The seasonal patterns of N-NO₃⁻ (Figure 2d-f), TDN (Figure 2j-l), pH

temperature (Figure 3a-c). The DO varied seasonally, with the peak in spring (Figure

272 2a-c). In contrast, N–NH₄⁺ concentrations were significantly lower in spring (Figure

273 **2g-i**).

274 3.2. Spatial Variation in Surface Water N₂O Concentration

Across all sampling sites in the reservoir, dissolved N₂O concentration in 275 November 2018, March 2019 and June 2019 ranged from 8.99 to 86.58, 6.11 to 276 110.37, and 7.33 to 261.27 nmol L^{-1} (Figure 4), respectively, with mean N₂O 277 concentration of 46.42 ± 2.03 , 58.96 ± 5.90 , and 43.96 ± 2.69 nmol L⁻¹ in the three 278 sampling campaigns. Across the three sampling campaigns, the mean N₂O 279 concentration were significantly greater in NRZ than in SRZ (p < 0.05, Two-way 280 ANOVAs; Table 1 and Figure S3a-c). Meanwhile, dissolved N₂O concentration also 281 showed remarkably differences betwee the wastewater loading area and the 282 non-wastewater loading area (p < 0.05; Table S4 and Figure S4a-c). Across the three 283 284 sampling campaigns, specifically, the mean N₂O concentration were significantly lower in Sector-N than the other sectors (p < 0.05; Figure 5a), and were generally 285 highest in Sector-R (Figure 5a). 286

287 3.3. Spatial Variation in Diffusive N₂O Fluxes Across the Water-Air Interface

Across all sampling sties, the N₂O fluxes ranged from 2.61 to 415.61, from -10.83 to 254.01, and from 2.04 to 711.50 μ g m⁻² h⁻¹ in November 2018, March 2019 and June 2019, respectively, with mean N₂O fluxes of 110.41±9.31, 78.48±6.27, and 153.12±8.61 μ g m⁻² h⁻¹ in the three sampling campaigns. Overall, the Wenwusha Reservoir acted as a constant source of N₂O to the atmosphere.

N₂O fluxes (Figure 6) showed a similar spatial variability to N₂O concentration
(Figure 4). Relatively higher N₂O fluxes were found in the NRZ of the Wenwusha

Reservoir. Across all sampling campaigns, the N₂O fluxes in the NRZ ranged from 6.87 to 711.50 μ g m⁻² h⁻¹, with an average of 153.12±8.61 μ g m⁻² h⁻¹, which was almost 3 times higher than those from the SRZ (*p*<0.001, Two-way ANOVAs; Table 1 and Figure S3d-f).

Significant differences in mean N₂O emission fluxes were also observed among the wastewater loading area and the non-wastewater loading area (p<0.05; Table S4 and Figures S4d-f). Across the three sampling campaigns, specifically, Sector-N had significantly lower mean N₂O fluxes than the other four wastewater loading sectors over the whole study period (Figure 5b), while Sector-R had relatively higher mean N₂O fluxes than other sectors (Figure 5b).

305 3.5. Water Quality Drivers of Variation in N₂O Concentrations and Fluxes

306 The simple regressions analysis demonstrated that he spatial variation of N₂O concentration and fluxes was positively related to the N-NO₃⁻ ($r^2 = 0.20-0.87$, 307 p < 0.01), N-NH₄⁺ ($r^2 = 0.17-0.42$, p < 0.01 or < 0.05) and TDN concentrations ($r^2 = 0.17-0.42$, p < 0.01 or < 0.05) 308 309 0.13–0.88, p<0.01) within each sampling occasion (Table S5, and Figures S5 and S6), respectively, and negatively correlated with that of salinity and DO (p < 0.05 or < 0.01; 310 311 Table S5). Principal component analysis (PCA) was also employed to explore the relationships among all these environmental variables over whole reservoir survey 312 during each sampling campaign (Figure 7a-c). The first component (PCA I), 313 explaining 93.1% (Figure 7a), 95.8% (Figure 7b) and 95.7 (Figure 7c) of the total 314 variances during the autumn, spring and summer surveys, respectively, was 315 significantly correlated with N–NO₃⁻, N–NH₄⁺ and TDN. The first component can be 316

considered as the influence of water pollution, reflecting the spatial variation. The 317 second component (PCA II), explaining 3.6%, 3.8% and 4.2% of the variances during 318 319 the autumn, spring and summer surveys, respectively, was correlated with the EC, salinity and DO. In addition, PCA was employed to explore the temporal correlation 320 321 between the N₂O concentration (or fluxes) and environmental variables. The first and second components together explained 98.2% of the total variance (Figure 7d). In 322 particular, the PCA I contributed 93.8% of the total variance and was correlated with 323 the $N-NO_3^-$, $N-NH_4^+$, TDN and temperature (Figure 7d). 324

325 3.6. Effects of Different Sampling Size on the Estimation of Whole-Reservoir N₂O
326 Fluxes

The results of Monte Carlo analysis showed that the accuracies of average diffusive N₂O fluxes across the whole reservoir increased considerably with the expansion of numbers of sampling sites included in flux estimations (Figure 8). For each sample size, the resampling process was repeated 10,000 times to create the boxplots of N₂O resampling means (Figure 8a) and the standard deviation of resampling means (Figure 8b).

In Figure 8a, the light grey, grey and dark grey shaded areas show the respective 70%, 95%, and 99% of the Highest Probability Density (HPD) ranges which indicate the variance of standard errors for approximate coverages of the resampling means from 10,000 simulations for each sample size. With the increase of sample sizes, the converging HPD ranges of possible resampling mean N_2O values indicated the improving estimation accuracy of N_2O flux. In addition, with the expansion of sample sizes, the biases of the resampling N_2O estimations declined, and the standard deviations of resampling means decreased exponentially (Figure 8b). Notably, the results of Monte Carlo analysis showed that increase of the sample size from 10 to 100 markedly reduced the relative standard deviation of mean diffusive N_2O flux from 21% to 1.2%.

The number of samples in Figure 8 could be used to determine an optimal 344 sample size to produce reasonable flux estimates. However, to achieve less than 5% of 345 error, more than 80 samples over the whole reservoir were still needed (ca. 15 samples 346 347 per km²), due to reservoir heterogeneity. If the sample sizes were less than 80, the uncertainty range of the resampling simulations increases because of missing some 348 measurements from the N₂O hotspots. Therefore, it is important to conduct high 349 350 spatial resolution sampling to include all N2O hotspots to improve the accuracy of flux estimates. 351

According to the 10,000 simulations of only 10 samples, a relative standard deviation of the estimated mean diffusive N₂O flux was 22.6% (Figure 8b). Despite a large range, these means had a median value of 107 μ mol m⁻² h⁻¹ (Figure 8a). The bias of the resampling median is small compared to the mean of all samples. Therefore, the high spatial resolution measurements from different transects can effectively reduce the estimation bias of N₂O flux in the current study.

Moreover, the standard deviations of resampled N₂O mean diffusive flux decreased dramatically from less than 10 μ mol m⁻² h⁻¹ for the sample size of 40 to 1.29 μ mol m⁻² h⁻¹ for the sample size of 100 (Figure 8b). The relative standard 361 deviation of mean diffusive flux reduced to only 1.2% for the sample size of 100, 362 indicating that the estimated whole-reservoir N_2O emissions approached the actual 363 mean, with much less variation. Increasing the sample size could effectively reduce 364 the estimation bias of the whole-reservoir N_2O emissions.

365 **4. Discussion**

366 4.1. Subtropical Coastal Reservoir are Atmospheric N₂O Sources

Results from our spatially intensive sampling showed that the average N₂O 367 emission flux from Wenwusha Reservoir were 107.0 µg m⁻² h⁻¹. Although, the average 368 N₂O fluxes in the Wenwusha Reservoir were smaller than those in some tropical 369 reservoirs (Table S6), for example, Tucuruiand reservoir, Samuel reservoir (Lima et 370 al., 2002), Petit Saut reservoir and Fortuna reservoir (Guérin et al., 2008), the N₂O 371 372 fluxes in our reservoir were one to two orders of magnitude higher than those in many reservoirs in subtropical (Liu et al., 2011a; Musenze et al., 2014; Shi et al., 2020; Yu 373 et al., 2018) and temperate regions (Cheng et al., 2019; Descloux et al., 2017; 374 375 Huttunen et al., 2002) (Table S6). The mean N₂O emission rates in our reservoir were also substantially higher than the median emission rate (7.2 μ g m⁻² h⁻¹) in global 376 lakes/reservoirs (Hu et al., 2016), and were approximately 2.5 times higher than the 377 average of China's reservoirs (42.3 μ g m⁻² h⁻¹) (Li et al., 2018). Compared with lakes 378 worldwide (Table S6), our reservoir had a larger N₂O emission fluxes. For example, 379 the mean N₂O emission rates in our reservoir was approximately 8.7 times higher than 380 the average of China's lakes (12.2 μ g m⁻² h⁻¹) (Li et al., 2018). In addition, the mean 381 N₂O emission from our reservoir was approximately 3.9 and 1.9 times higher than 382

emissions from *Cyperus malaccensis* marsh (Wang et al., 2018) and aquaculture ponds (Yang et al., 2020) in the same subtropical coastal region. Futhermore, high N₂O emission fluxes has been reported in other subtropical coastal reservoirs studies (e.g., Cheng et al., 2014; Zhang, 2012). These results together indicate that subtropical costal reservoirs could be important sources of atmospheric N₂O and therefore more attention is needed.

389 4.2. External Sewage Input Affects the Spatial Pattern of N₂O Emission in the
390 Reservoir

391 There are some studies on the small-scale spatial variations in N₂O flux across the water-air interface in inland aquatic ecosystems (e.g., reservoirs, lakes) (Cheng et 392 al., 2019; Musenze et al., 2014; Zhao et al., 2013; Xiao et al., 2019a). However, such 393 394 knowledge is still lacking in coastal reservoirs. In the current study, our intensive sampling in multiple sites shed light on the marked spatial variation in N₂O emissions 395 in Wenwusha Reservoirin (Figure 6), and the coefficients of variation changed from 396 397 81% to 110% in the three sampling campaigns. Notably, the wastewater loading area only took up 30% of the whole reservoir area, but it contributed approximately 60% 398 of N₂O emissions in the whole reservoir. 399

Some scholars have found that the microbial processes of N₂O production can be promoted by large provision of of N substrate, subsequently increasing water N₂O concentration and emission to atmosphere (Herrman et al., 2008; Huttunen et al., 2003; Liu et al., 2011a; Xiao et al., 2019a, 2019b; Yu et al., 2013; Zhao et al., 2014). In this study, some environmental parameters are relative stable in the whole reservoir, for

example, air temperature, atmospheric pressure, solar radiation, and wind speed 405 (wind-dependent gas transfer coefficient), but some parameters had marked spatial 406 407 variations, such as the concentrations of N-NH₄⁺, N-NO₃⁻, and TDN (Figure 2). Located in a catchment with intensive human activity, the Wenwusha Reservoir 408 409 catchment is heavily affected by anthropogenic activities, particularly wastewater 410 discharge from domestic and industrial sources, and drainage from mariculture ponds, (Figure 1). Results in our reservoir indicated the the highest average concentrations of 411 N-NH₄⁺, N-NO₃⁻, and TDN from the river input sector, suggesting the important effect 412 413 of the upstream area in providing nitrogen to the waterbody. Different from other wastewater loading sectors, non-wastewater loading sector had much lower N-NH4⁺, 414 N-NO₃⁻, and TDN concentrations and obviously higher DO level (Table S3), 415 416 suggesting a improved water quality without the impact of sewage input. Therefore, the changing levels of sewage discharge was one of the primary factors controlling the 417 large spatial variation in N₂O emissions (Figure 6). 418

419 The changes of N₂O fluxes between the two reservoir zones (Figure S3) further confirmed the role of sewage discharge in controlling N2O emissions from the 420 421 Wenwusha Reservoir. During our study period, N₂O concentrations (Figure 4 and Figure S3a) and emission fluxes (Figure 6 and Figure S3a) in the NRZ were much 422 higher than those in the SRZ. This is likely due to that the markedly higher 423 concentrations of N-NH4⁺, N-NO₃⁻, and TDN (Figure 2) enhanced N₂O production in 424 NRZ, as a consequence of more input from wastewater (Figure 1). The relationship 425 between sewage input, N substrates, and N₂O emission has been found in lakes (e.g., 426

427 Xiao et al., 2019a), coastal river network (Yu et al., 2013), and inland reservoirs (e.g.,

He et al., 2017; Liu et al., 2011a, 2017). In the current study, N₂O production data is unavailable. However, the significantly positive relationships between the N₂O concentration (NO₂ emission fluxes) and N concentrations in Wenwusha reservoir (p<0.05 or <0.01; Table S4 and Figures 5-7) confirmed the above explanation.

The dissolved N₂O concentrations in the wastewater of municipal sewage 432 drainage channels, aquaculture ponds, and rivers adjacent to the reservoir were about 433 3-5 times higher than those in the reservoir surface water, and this caused a direct 434 435 input of dissolved N₂O into the Wenwusha Reservoir, with a consequence of steeper gradients of N2O concentrations between the surface water and atmosphere and 436 large N₂O diffusive emissions. Despite some environmental protection measures, a 437 438 large amount of sewage was still discharged into the reservoir. This external N2O input can support N₂O emissions continuously from water surface into air, even lack 439 of internal N₂O production inside the water ecosystem. Certainly, the contributions of 440 441 sewage discharge to the direct input of N₂O into the waterbody should not be overlooked. Effective measures are urgently needed to reduce and finally stop the 442 443 discharge of sewage into the reservoir.

444 *4.3.* Role of Salinity in the Spatial Variation in Reservoir N₂O Emission

Except for N availability, the spatial variation in N_2O concentrations and fluxes in the Wenwusha Reservoir may be influenced by the difference in salinity. Salinity controls the mineralization rate of N through its effect on microbial activity, enzyme, and metabolism, and consequently regulates the rates of N_2O production,

449	consumption, and emission into the atmosphere. Some studies suggested that high
450	salinity can reduce microbial diversity (Francis et al., 2003; Li et al., 2020; Moin et al.,
451	2009; Mosier and Francis, 2008) and activity (Wang et al., 2010), especially the
452	ammonia-oxidizing bacteria (AOBs), with the consequence of reducing sediment /
453	water N ₂ O production (Rysgaard et al., 1999; Wang et al., 2018). The negative
454	relationship between salinity and N2O concentrations (flux) has been reported in
455	coastal wetlands and aquatic ecosystems (Liu et al., 2015; Sun et al., 2015; Wang et
456	al., 2018; Welti et al., 2017). In the present study, salinity in the surface water showed
457	remarkable spatial variations (Figure 3j-l), with the NRZ exhibiting significantly
458	lower salinity than the SRZ ($p < 0.01$; Figure S3c), as a result of freshwater dilution
459	caused by the river runoff (Figure 1). The salinity (Figure 3j-l) and N ₂ O concentration
460	(or flux) (Figure 4 and 6) exhibited opposite spatial patterns in the reservoir (p <0.01;
461	Table S5), indicating that salinity might be an important environmental factor
462	influencing N ₂ O dynamics between the two reservoir zones. Yet, salinity was likely
463	not a dominant factor in governing the spatial variations of N ₂ O emission fluxes in
464	each zone, since the spatial heterogeneity of water salinity within each zone was very
465	low (Figure 3j-1). Meanwhile, the substantial differences in salinity beween two
466	reservoir zones also provide additional evidence for that the large contribution of
467	external N loading to the spatial variation in N2O fluxes within Wenwusha Reservoir.
468	Further studies merit to explore the exact impacts of salinity on N ₂ O production and
469	emission.

470 4.4. Temporal Variation in N₂O Emission Fluxes Across the Coastal Reservoir

471	In this study, the N ₂ O flux exhibited clear seasonal variation (p <0.01; Table 1),
472	with significant higher values in summer (Figure 5). Similar seasonal pattern was also
473	observed in inland freshwaters (Musenze et al., 2014; Xiao et al., 2019a, 2019b; Zhu
474	et al., 2013). Temperature is generally considered as the most important factor driving
475	N ₂ O emission temporal patterns due to its stimulation of microbial activity related to
476	N ₂ O production (Beaulieu et al. 2010; Harrison and Matson, 2003; Hinshaw and
477	Dahlgren 2013). Larger N ₂ O fluxes in summer than autumn and spring (Figure 5)
478	coincided with higher warmer temperatures in summer (Figure S1a) across the whole
479	reservoir. However, an interesting outcome was that, although the water temperature
480	was higher in spring than autumn (Figure S1a), the N2O concentration and flux
481	(Figures 3 and 5) showed an opposite patterns with water temperature. This was
482	probably related to the disturbance of precipitation. In our study period, several
483	continuous rain events occurred during the spring surveys. The large precipitation not
484	only dramatically increased water depth in the reservoir, but also increased inflow of
485	water and soil drained into the reservoir by rain falling on surrounding land,
486	ultimately causing a dilution effect on many of the reservoir's biological and chemical
487	parameters. Thus, instead of higher temperature, the dilution effect from large rainfall
488	and river flows may be responsible for the lower N ₂ O concentration in spring than in
489	autumn. Similar results were reported by He et al. (2017) and Outram and Hiscock
490	(2012). In addition, annual drainage is a typical management activity that is practiced
491	in autumn period after mariculture harvest, and it is a way to export aquaculture
492	effluent. Thus, the N ₂ O concentrations and emission fluxes in autumn were larger than

those in spring also might be related to the aquaculture effluent discharge, which provides a large amount of N substrate for reservoir's N₂O production. This hypothesis was supported by the evidence that N₂O concentrations and emission fluxes showed significantly positive relationships with N–NO₃⁻, N–NH₄⁺ and TDN (p<0.01; Table S5), respectively. These results together indicated that the seasonal variations in the N₂O concentrations and emission fluxes in the Wenwusha Reservoir were regulated synthetically by temperature, precipitation and external N input.

500 4.5. Implications of Spatially Resolved Measurements of Reservoir N₂O Emission

501 In the past twenty years, scholars have gradually realized the large N₂O emission from reservoirs to atmosphere (e.g., Deemer et al., 2016; Descloux et al., 2017; 502 503 Guérin et al., 2008; Li et al., 2018; Maavara et al., 2019; Wang et al., 2017). However, 504 the accurate contribution of reservoirs to the global N₂O budget remains highly uncertain. Our results show that subtropical costal reservoirs are important 505 atmospheric N₂O sources with substantive spatial variability. The large spatial 506 507 variability in N₂O fluxes within the reservoir implies a large uncertainty of the whole reservoir estimate of N₂O fluxes provided by earlier studies that were conventionally 508 based on single site or limited number of site measurements. The results of Monte 509 Carlo analysis further showed that increasing the sample size from 10 to 100 510 significantly reduced the relative standard deviation of mean diffusive N₂O flux from 511 22.6% to 1.6% (Figure 8). In the process of extrapolating small-scale N₂O fluxes to 512 513 the whole reservoir, therefore, our results highlight the importance of conducting high spatial resolution sampling (>15 samples per km^2) to minimize the bias. 514

515	Our high spatial resolution sampling data show significant larger N ₂ O emissions
516	from the wastewater loading area, indicating the important role of wastewater input
517	for promoting N ₂ O emissions from reservoirs. With the world entering anthropocene,
518	human activities, particularly wastewater discharge, have severely increased the input
519	of anthropogenic nutrients to aquatic ecosystems, particularly in developing countries
520	with poor wastewater treatment system (Hosen et al., 2014; Williams et al., 2016;
521	Yang et al., 2013; Yu et al., 2013), but insufficient attention has been given to
522	anthropogenic activities as an accelerator influencing the N ₂ O and other greenhouse
523	gas emissions from water (He et al., 2017; Wang et al., 2017; Xiao et al., 2019a; Xing
524	et al., 2006; Yang et al., 2008). Clearly, strengthening environmental protection,
525	particularly wastewater treatment and discharge, is the key to mitigate water pollution
526	and greenhouse emission in aquatic environment (Yang, 2014; Yang et al., 2015b).

527 This study, to the best of our knowledge, represents the first attempt to quantify 528 N_2O emission fluxes from costal reservoir at such a high spatial resolution. Overall, 529 the results provide a valuable data source for model development and validation to 530 predict N_2O emission fluxes from costal reservoirs suffering from intensive human 531 disturbance.

532 4.6. Limitations and Future Research

533 Similar to most studies, there are limitations in the current study. First, diurnal 534 variations in N₂O flux have been reported in inland freshwaters (Baulch et al., 2012; 535 Laursen and Seitzinger, 2004; Wu et al., 2018; Xiao et al., 2019a; Yang et al., 2011), 536 and some researches found larger fluxes during nighttime compared with daytime

(Baulch et al., 2012; Wu et al., 2018; Yang et al., 2011). However, the day-night 537 pattern of N₂O fluxes in the Wenwusha Reservoir is unknown, which may lead to 538 539 estimate uncertainty of seasonal or annual N₂O budget. Therefore, future long-term observations along with diurnal temporal scales can further reduce the estimate bias of 540 541 N₂O emission budget. Moreover, thermal stratification is a phenomenon frequently happening in reservoirs, particularly in deep reservoir (Straskraba, 1999; Thornton, 542 1990). The stratification could create barriers to the exchange of material (e.g., 543 nutrient and DO) between the epilimnion and the hypolimnion (Hayes et al., 2107; Yu 544 545 et al., 2015), ultimately resulting in a change of the main pathways of N₂O production in the water profile (Beaulieu et al., 2015; Liang et al., 2019; Salk et al., 2016) and the 546 subsequent emission to the atmosphere. However, the vertical pattern of N₂O dynamic 547 548 within reservoir in the current study is unknown, which limits the understanding of N₂O production and spatial patterns. In the future work, therefore, there is an urgent 549 need for utilizing advancing technologies (e.g., isotopic tracing) to investigate the 550 vertical profile of N₂O dynamic in coastal reservoirs. Furthermore, this study 551 examined N₂O concentrations and fluxes, and basic environmental factors in situ, 552 while the contributions from other N₂O production pathways, and microbial 553 abundance and activity (nitrifier, denitrifier, and ammonifier) are unclear. Thus, the 554 future detailed investigation of the more production pathways and microbial 555 mechanisms of N₂O in coastal reservoirs will be helpful for understanding the 556 mysterious N cycle in aquatic ecosystems. 557

558 **5. Conclusions**

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559	We estimated N ₂ O emissions from Wenwusha Reservoir based on high spatial
560	resolution data. Total surface water diffusive N2O fluxes was estimated to be 4.8 Mg
561	yr ⁻¹ , with an annual average CH ₄ flux of 107.0 μ g m ⁻² h ⁻¹ . Fluxes in N ₂ O were much
562	greater in sectors in which sewage is discharged to the reservoir, due to greater inputs
563	of anthropogenic N. As a result of high levels of nutrient and organic matter loading,
564	areas of wastewater loading (comprising 30% of the reservoir area) acted as potential
565	N ₂ O emission hotspots, and made a disproportionate contribution (60%) to
566	whole-reservoir N_2O emissions. Our results illustrate that coastal reservoirs are an
567	important source of atmospheric N2O, where spatial variation in emissions is driven
568	by anthropogenic activity. To produce representative, accurate estimates of N_2O
569	emissions from coastal reservoirs suffering anthropogenic disturbance, it is essential
570	they are based on highly spatially resolved data.

571 **Declaration of interest**

572 None

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584 **References**

- Barnard, R., Leadley, P., Hungate, B.A., 2005. Global change, nitrification, and denitrification: a
 review. Global Biogeochem. Cy. 19, 2282–2298. https://doi.org/10.1029/2004GB002282
- Baulch, H.M., Dillon, P.J., Maranger, R., Venkiteswaran, J.J., Wilson, H.F., Schiff, S.L., 2012.
 Night and day: short term variation in nitrogen chemistry and nitrous oxide emissions from
 streams. Freshwater Biol. 57, 509–525. https://doi.org/10.1111/j.1365-2427.2011.02720.x
- Beaulieu, J.J., Nietch, C.T., Young, J.L., 2015. Controls on nitrous oxide production and
 consumption in reservoirs of the Ohio River Basin. J. Geophys. Res. Biogeosci. 120(10),
 1995–2010. https://doi.org/10.1002/2015JG002941
- Beaulieu, J.J., Shuster, W.D., Rebholz, J.A., 2010. Nitrous oxide emissions from a large,
 impounded river: the Ohio River. Environ. Sci. Technol. 44, 7527–7533.
 https://doi.org/10.1021/es1016735
- 596 Beaulieu, J.J., Tank, J.L., Hamilton, S.K., Wollheim, W.M., Hall Jr., R.O., Mulholland, P.J.,
- 597 Peterson, B.J., Ashkenas, L.R., Cooper, L.W., Dahm, C.N., Dodds, W.K., Grimm, N.B.,
- 598 Johnson, S.L., McDowell, W.H., Poole, G.C., Valett, H.M., Arango, C.P., Bernot, M.J., Burgin,
- 599 A.J., Crenshaw, C.L., Helton, A.M., Johnson, L.T., O'Brien, J.M., Potter, J.D., Sheibley, R.W.,
- 600 Sobota, D.J., Thomas, S.M., 2011. Nitrous oxide emission from denitrification in stream and
- 601 river networks. Proc. Natl. Acad. Sci. U.S.A. 108, 214–219.
 602 https://doi.org/10.1073/pnas.1011464108
- Borges, A.V., Speeckaert, G., Champenois, W., Scranton, M.I., Gypens, N., 2018. Productivity and
 temperature as drivers of seasonal and spatial variations of dissolved methane in the Southern
 Bight of the North Sea. Ecosystems 21(4), 583–599.
 https://doi.org/10.1007/s10021-017-0171-7.
- Brase, L., Bange, H.W., Lendt, R., Sanders, T., Dähnke, K., 2017. High resolution measurements
 of nitrous oxide (N₂O) in the Elbe estuary. Front. Mar. Sci. 4, 162.
 https://doi.org/10.3389/fmars.2017.00162

- 610 Cebron, A., Garnier, J., Billen, G., 2005. Nitrous oxide production and nitrification kinetics by
 611 natural bacterial communities of the lower Seine river (France). Aquat. Microb. Ecol. 41,
 612 25–38. https://doi.org/10.3354/ame041025
- 613 Chen, N.W., Chen, Z.H., Wu, Y.Q., Hu, A.Y., 2014. Understanding gaseous nitrogen removal
 614 through direct measurement of dissolved N₂ and N₂O in a subtropical river-reservoir system.
- 615 Ecol. Eng. 70, 56–67. https://doi.org/10.1016/j.ecoleng.2014.04.017
- 616 Cheng, F., Zhang, H.M., Zhang, G.L., Liu, S.M., Song, G.D., Du, G.X., 2019. Distribution and
 617 emission of N₂O in the largest river-reservoir system along the Yellow River. Sci. Total
 618 Environ. 666, 1209–1219. https://doi.org/10.1016/j.scitotenv.2019.02.277
- Cohen, Y., Gordon, L.I., 1979. Nitrous oxide production in the ocean. J. Geophys. Res. Oceans 84,
 347–353. https://doi.org/10.1029/JC084iC01p00347
- 621 Cole, J.J., Caraco, N.F., 1998. Atmospheric exchange of carbon dioxide in a low-wind oligotrophic
- 622 lake measured by the addition of SF₆. Limnol. Oceanogr. 43, 647–656.
 623 https://doi.org/10.4319/lo.1998.43.4.0647
- Cooper, R.J., Wexler, S.K., Adams, C.A., Hiscock, K.M., 2017. Hydrogeological controls on
 regional-scale indirect nitrous oxide emission factors for rivers. Environ. Sci. Technol. 51,
 10440–10448. https://doi.org/10.1021/acs.est.7b02135
- 627 Crusius, J., Wanninkhof, R., 2003. Gas transfer velocities measured at low wind speed over a lake.
 628 Limnol. Oceanogr. 48(3), 1010-1017.
- 629 Davidson, E.A., 2009. The contribution of manure and fertilizer nitrogen to atmospheric nitrous
- 630 oxide since 1860. Nat. Geosci. 2, 659-662. https://doi.org/10.1038/ngeo608
- 631 Deemer, B.R., Harrison, J.A., Li, S.Y., Beaulieu, J.J., Delsontro, T., Barros, N., Bezerra-Neto, J.F.,
- Powers, S.M., Santos, M.A.D., Vonk, J.A., 2016. Greenhouse gas emissions from reservoir
 water surfaces: a new global synthesis. BioScience 66, 949–964.
 https://doi.org/10.1093/biosci/biw117
- Deemer, B.R., Harrison, J.A., Whitling, E.W., 2011. Microbial dinitrogen and nitrous oxide
 production in a small eutrophic reservoir: an in situ approach to quantifying hypolimnetic
- 637 process rates. Limnol. Oceanogr. 56:1189-1199. https://doi.org/10.4319/lo.2011.56.4.1189
- 638 De Klein, C., et al., 2006. N₂O emissions from managed soils, and CO₂ emissions from lime and
- 639 urea application. IPCC Guidelines for National Greenhouse Gas Inventories, Prepared by the
- 640 National Greenhouse Gas Inventories Programme, 4, pp. 1–54.

- 641 Descloux, S., Chanudet, V., Serça, D., Guérin, F., 2017. Methane and nitrous oxide annual
 642 emissions from an old eutrophic temperate reservoir. Sci. Total Environ. 598, 959–972.
 643 https://doi.org/10.1016/j.scitotenv.2017.04.066
- Francis, C.A., O'Mullan, G.D., Ward, B.B., 2003. Diversity of ammonia monooxygenase (*amoA*)
 genes across environmental gradients in Chesapeake Bay sediments. Geobiology 1, 129–140.
 https://doi.org/10.1046/j.1472-4669.2003.00010.x
- Fu, C., Lee, X., Griffis, T.J., Baker, J.M., Turner, P.A., 2018. A modeling study of direct and
 indirect N₂O emissions from a representative catchment in the U.S. Corn Belt. Water Resour.
- 649 Res. 54(5), 3632–3653. https://doi.org/10.1029/2017WR022108
- Garnier, J., Cébron, A., Tallec, G., Billen, G., Sebilo, M., Martinez, A., 2006. Nitrogen behaviour
 and nitrous oxide emission in the tidal Seine River Estuary (France) as influenced by human
 activities in the upstream watershed. Biogeochemistry 77, 305–326.
 https://doi.org/10.1007/s10533-005-0544-4
- Gruber, N., Galloway, J.N., 2008. An Earth-system view of the global nitrogen cycle. Nature 451,
 293–296. https://doi.org/10.1038/nature06592
- Guérin, F., Abril, G., Tremblay, A., Delmas, R., 2008. Nitrous oxide emissions from tropical
 hydroelectric reservoirs. Geophys. Res. Lett. 35(6), L06404.
 https://doi.org/10.1029/2007GL033057
- Hama-Aziz, Z.Q., Hiscock, K.M., Cooper, R.J., 2017. Indirect nitrous oxide emission factors for
 agricultural field drains and headwater streams. Environ. Sci. Technol. 51, 301–307.
 https://doi.org/10.1021/acs.est.6b05094
- Hampel, J.J., McCarthy, M., Gardner, W., Zhang, L., Xu, H., Zhu, G., Newell, S., 2018.
 Nitrification and ammonium dynamics in Taihu Lake, China: Seasonal competition for
 ammonium between nitrifiers and cyanobacteria. Biogeosciences 15, 733–748.
 https://doi.org/10.5194/bg-15-733-2018
- Harrison, J., Matson, P., 2003. Patterns and controls of nitrous oxide emissions from waters
 draining a subtropical agricultural valley. Glob. Biogeochem. Cy 17.
 https://doi.org/10.1029/2002GB001991
- Hayes, N.M., Deemer, B.R., Corman, J.R., Razavi, N.R., Strock, K.E., 2017. Key differences
 between lakes and reservoirs modify climate signals: A case for a new conceptual model.
- 671 Limnol. Oceanogr. Lett. 2, 47–62. https://doi.org/10.1002/lol2.10036

- He, Y.X., Wang, X.F., Chen, H., Yuan, X.Z., Wu, N., Zhang, Y.W., Yue, J.S., Zhang, Q.Y., Diao,
- Y.B., Zhou, L.L., 2017. Effect of watershed urbanization on N₂O emissions from the
 Chongqing metropolitan river network, China. Atmos. Environ. 171, 70–81.
 https://doi.org/10.1016/j.atmosenv.2017.09.043
- Herrman, K.S., Bouchard, V., Moore, R.H., 2008. Factors affecting denitrification in agricultural
 headwater streams in Northeast Ohio, USA. Hydrobiologia 598, 305–314.
 https://doi.org/10.1007/s10750-007-9164-4
- Hinshaw, S. E., and R. A. Dahlgren. 2013. Dissolved nitrous oxide concentrations and fluxes from
 the eutrophic San Joaquin River, California. Environ. Sci. Technol. 47, 1313–1322.
 https://doi.org/10.1021/es301373h
- Hirota, M., Senga, Y., Seike, Y., Nohara, S., Kunii, H., 2007b. Fluxes of carbon dioxide, methane
 and nitrous oxide in two contrastive fringing zones of coastal lagoon, Lake Nakaumi, Japan.
 Chemosphere 68 (3), 597–603. https://doi.org/10.1016/j.chemosphere.2007.01.002
- Hosen, J.D., McDonough, O.T., Febria, C.M., Palmer, M.A., 2014. Dissolved organic matter
 quality and bioavailability changes across an urbanization gradient in headwater streams.

687 Environ. Sci. Technol. 48, 7817–7824. https://doi.org/10.1021/es501422z

- Hu, M., Chen, D., Dahlgren, R.A., 2016. Modeling nitrous oxide emission from rivers: a global
 assessment. Glob. Chang. Biol. 22, 3566-3582. https://doi.org/10.1111/gcb.13351
- 690 Huttunen, J.T., Alm, J., Liikanen, A., Juutinen, S., Larmola, T., Hammar, T., Silvola, J.,
- 691 Martikainen, P.J., 2003. Fluxes of methane, carbon dioxide and nitrous oxide in boreal lakes
- and potential anthropogenic effects on the aquatic greenhouse gas emissions. Chemosphere 52,
- 693 609-621. https://doi.org/10.1016/S0045-6535(03)00243-1
- Huttunen, J.T., Vaisanen, T.S., Hellsten, S.K., Heikkinen, M., Nykanen, H., Jungner, H., Niskanen,
- A., Virtanen, M.O., Lindqvist, O.V., Nenonen, O.S., Martikainen, P.J., 2002. Fluxes of CH₄,
- 696 CO₂, and N₂O in hydroelectric reservoirs Lokka and Porttipahta in the northern boreal zone in
- 697 Finland. Glob Biogeochem Cy. 16(1), 1003. https://doi.org/10.1029/2000GB001316
- Hinshaw, S.E., Dahlgren, R.A., 2013. Dissolved nitrous oxide concentrations and fluxes from the
 eutrophic San Joaquin River, California. Environ. Sci. Technol. 47, 1313–1322.
 https://doi.org/10.1021/es301373h

- Hosen, J.D., McDonough, O.T., Febria, C.M., Palmer, M.A., 2014. Dissolved organic matter
 quality and bioavailability changes across an urbanization gradient in headwater streams.
 Environ. Sci. Technol. 48, 7817-7824. https://doi.org/10.1021/es501422z
- 704 IPCC, 2013. In: T. F. Stocker, et al. (Eds.), Climate change 2013: The physical science basis. 705 Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental 706 Panel on Climate Change. Cambridge: Cambridge University Press. doi: 707 10.1017/CBO9781107415324
- Kester, R.A., De Boer, W., Laanbroek, H.J., 1997. Production of NO and N₂O by pure cultures of
 nitrifying and denitrifying bacteria during changes in aeration. Appl. Environ. Microb. 63,
 3872–3877.
- Laursen, A.E., Seitzinger, S.P., 2004. Diurnal patterns of denitrification, oxygen consumption and
 nitrous oxide production in rivers measured at the whole-reach scale. Freshw. Biol. 49,
 1448–1458. https://doi.org/10.1111/j.1365-2427.2004.01280.x
- Li, S.Y., Bush, R.T., Santos, I.R., Zhang, Q.F., Song, K.S., Mao, R., Wen, Z.D., Lu, X.X., 2018.
 Large greenhouse gases emissions from China's lakes and reservoirs. Water Res. 147, 13–24.

716 https://doi.org/10.1016/j.watres.2018.09.053

- Li, Y.X., Zhang, L.J., Xue, L., Fan, W.H., Liu, F.W., Yang, H., 2020. Spatial variation in aragonite
 saturation state and the influencing factors in Jiaozhou Bay, China. Water 12(3), 825.
 https://doi.org/10.3390/w12030825
- 720 Liang, X., Xing, T., Li, J.X., Wang, B.L., Wang, F.S., He, C.Q., Hou, L.J., Li, S.L., 2019. Control
- of the hydraulic load on nitrous oxide emissions from cascade reservoirs. Environ. Sci.
 Technol. 53(20), 11745–11754. https://doi.org/10.1021/acs.est.9b03438
- 723 Lima, I.B.T., Victoria, R.L., Novo, E.M.L.M., Feigl, B.J., Ballester, M.V.R., Ometto, J.P., 2002. 724 Methane, carbon dioxide and nitrous oxide emissions from two Amazonian reservoirs during 725 high water table. Verh. Internat. Verein. Limnol. 28, 438-442. 726 https://doi.org/10.1080/03680770.2001.11902620
- 727 Liu, X.L., Li, S.L., Wang, Z.L., Han, G.L., Li, J., Wang, B.L., Wang, F.S., Bai, L., 2017. Nitrous
- oxide (N₂O) emissions from a mesotrophic reservoir on the Wujiang River, southwest China.
- 729 Acta Geochim 36(4), 667–679. https://doi.org/10.1007/s11631-017-0172-4

- Liu, X.L., Liu, C.Q., Li, S.L., Wang, F.S., Wang, B.L., Wang, Z.L., 2011a. Spatiotemporal
 variations of nitrous oxide (N₂O) emissions from two reservoirs in SW China. Atmos. Environ.
 45 (31), 5458–5468. https://doi.org/10.1016/j.atmosenv.2011.06.074.
- Liu, X.L., Bai, L., Wang, Z.L., Li, J., Yue, F.J., Li, S.L., 2015. Nitrous oxide emissions from river
 network with variable nitrogen loading in Tianjin, China. J. Geochem. Explor. 157, 153–161.
 https://doi.org/10.1016/j.gexplo.2015.06.009.
- Liu, X.L., Li, S.L., Wang, Z.L., Han, J.L., Li, J., Wang, B.L., Wang, F.S., Bai, L., 2017. Nitrous
 oxide (N₂O) emissions from a mesotrophic reservoir on the Wujiang River, Southwest China.
- 738 Acta Geochim. 36 (4), 667–679. https://doi.org/10.1007/s11631-017-0172-4
- 739 Liu, Y.S., Zhu, R.B., Ma, D.W., Xu, H., Luo, Y.H., Huang, T., Sun, L.G., 2011b. Temporal and
- spatial variations of nitrous oxide fluxes from the littoral zones of three alga-rich lakes in
 coastal Antarctica. Atmos. Environ. 45, 1464–1475.
 https://doi.org/10.1016/j.atmosenv.2010.12.017
- Maavara, T., Lauerwald, R., Laruelle, G.G., Akbarzadeh, Z., Bouskill, N.J., Van Cappellen, P.,
 Regnier, P., 2019. Nitrous oxide emissions from inland waters: Are IPCC estimates too high?
 Global Change Biol. 25, 473–488. https://doi.org/10.1111/gcb.14504
- Mengis, M., Gachter, R., Wehrli, B., 1996. Nitrous oxide emissions to the atmosphere from an
 artificially oxygenated lake. Limnol. Oceanogr. 41, 548–553.
 https://doi.org/10.4319/lo.1996.41.3.0548.
- Mengis, M., Gachter, R., Wehrli, B., 1997. Sources and sinks of nitrous oxide (N₂O) in deep lakes.
 Biogeochemistry 38, 281–301.
- Moin, N.S., Nelson, K.A., Bush, A., Bush, A., Bernhard, A.E., 2009. Distribution and diversity of
 archaeal and bacterial ammonia oxidizers in salt marsh sediments. Appl. Environ. Microbiol.
- 753 7 (23), 7461–7468. https://doi.org/10.1128/AEM.01001-09
- Mosier, A.C., Francis, C.A., 2008. Relative abundance and diversity of ammonia-oxidizing
 archaea and bacteria in the San Francisco Bay estuary. Environ. Microbiol. 10(11),
 3002–3016. https://doi.org/10.1111/j.1462-2920.2008.01764.x
- 757 Mosier, A., Kroeze, C., Nevison, C., Oenema, O., Seitzinger, S., van Cleemput, O., 1998. Closing
- the global N_2O budget: nitrous oxide emissions through the agricultural nitrogen cycle.
- 759 Nutrient Cycl. Agroecosyst. 52, 225-248.

- 760 Mulholland, P.J., Helton, A., Poole, G.C., Hall, R.O., Jr., Hamilton, S.K., Peterson, B.J., Tank, J.L.,
- 761 Ashkenas, L.R., Cooper, L.W., Dahm, C.N., Dodds, W.K., Findlay, S.E.G., Gregory, S.V.,
- 762 Grimm, N.B., Johnson, S.L., McDowell, W.H., Meyer, J.L., Valett, H.M., Webster, J.R.,
- 763 Arango, C.P., Beaulieu, J.J., Bernot, M.J., Burgin, A.J., Crenshaw, C.L., Johnson, L.T.,
- 764 Niederlehner, B.R., O' Brien, J.M., Potter, J.D., Sheibley, R.W., Sobota, D.J., Thomas, S.M.
- 765 2008. Stream denitrification across biomes and its response to anthropogenic nitrate loading.
- 766 Nature 452, 202–206. https://doi.org/10.1038/nature06686
- Murray, R.H., Erler, D.V., Eyre, B.D., 2015. Nitrous oxide fluxes in estuarine environments:
 response to global change. Global Change Biol. 21(9), 3219–3245.
 https://doi.org/10.1111/gcb.12923
- Musenze, R.S., Grinham, A., Werner, U., Gale, D., Sturm, K., Udy, J., Yuan, Z.G., 2014. Assessing
 the spatial and temporal variability of diffusive methane and nitrous oxide emissions from
 subtropical freshwater reservoirs. Environ. Sci. Technol. 48, 14499–14507.
 https://doi.org/10.1021/es505324h
- Najjar, R.G., 1992. "Marine biogeochemistry," in *Climate System Modeling*, ed K.E. Trenberth
 (Cambridge: Cambridge University Press), 241–280.
- Nevison, C., Butler, J.H., and Elkins, J., 2003. Global distribution of N2O and the ΔN₂O-AOU
 yield in the subsurface ocean. Global Biogeochem. Cycles 17(4), 1119.
 https://doi.org/10.1029/2003GB002068
- 779 Outram, F.N., Hiscock, K.M., 2012. Indirect nitrous oxide emissions from surface water bodies in
- a lowland arable catchment: a significant contribution to agricultural greenhouse gas budgets?
- 781 Environ. Sci. Technol. 46, 8156–8163. https://doi.org/10.1021/es3012244
- Ravishankara, A.R., Daniel, J.S., Portmann, R.W., 2009. Nitrous oxide (N₂O): The dominant
 ozone-depleting substance emitted in the 21st century. Science 326, 123–125.
 https://doi.org/10.1126/science.1176985
- Rosamond, M.S., Thuss, S.J., Schiff, S.L., 2012. Dependence of riverine nitrous oxide emissions
 on dissolved oxygen levels. Nat. Geosci. 5, 715-718. https://doi.org/10.1038/ngeo1556
- 787 Rosamond, M.S., Thuss, S.J., Schiff, S.L., Elgood, R.J., 2011. Coupled cycles of dissolved oxygen
- and nitrous oxide in rivers along a trophic gradient in Southern Ontario, Canada. J. Environ.
- 789 Qual. 40, 256–270. https://doi.org/10.2134/jeq2010.0009

- Rysgaard, S., Thastum, P., Dalsgaard, T., Christensen, P.B., Sloth, N.P., 1999. Effects of salinity on
 NH₄⁺ adsorption capacity, nitri fication, and denitri fication in Danish estuarine sediments.
 Estuar. Coasts 22, 21–30.
- 793 Saikawa, E., Prinn, R.G., Dlugokencky, E., Ishijima, K., Dutton, G.S., Hall, B.D., Langenfelds, R.,
- 794 Tohjima, Y., Machida, T., Manizza, M., Rigby, M., O'Doherty, S., Patra, P.K., Harth, C.M.,
- 795 Weiss, R.F., Krummel, P.B., van der Schoot, M., Fraser, P.J., Steele, L.P., Aoki, S.,
- 796 Nakazawa, T., Elkins, J.W., 2014. Global and regional emissions estimates for N₂O. Atmos.
- 797 Chem. Phys. 14, 4617–4641. https://doi.org/10.5194/acp-14-4617-2014
- Salk, K.R., Ostrom, P.H., Biddanda, B.A., Weinke, A.D., Kendall, S.T., Ostrom, N.E., 2016.
 Ecosystem metabolism and greenhouse gas production in a mesotrophic northern temperate
 lake experiencing seasonal hypoxia. Biogeochemistry 131, 303–319.
 https://doi.org/10.1007/s10533-016-0280-y
- 802 Sasaki, Y., Koba, K., Yamamoto, M., Makabe, A., Ueno, Y., Nakagawa, M., Toyoda, S., Yoshida, 803 N., Yoh, M., 2011. Biogeochemistry of nitrous oxide in Lake Kizaki, Japan, elucidated by 804 G04030. nitrous oxide isotopomer analysis. J. Geophys. Res. 116, 805 https://doi.org/10.1029/2010jg001589
- Shaaban, M., Wu, Y., Khalid, M., Peng, Q., Xu, X., Wu, L., Younas, A., Bashir, S., Mo, Y., Lin,
 S., Zafar-ul-Hye, M., Abid, M., Hu, R., 2018. Reduction in soil N₂O emissions by pH
 manipulation and enhanced nosZ gene transcription under different water regimes. Environ.
 Pollut. 235, 625–631. https://doi.org/10.1016/j.envpol.2017.12.066
- 810 Shi, W.Q., Chen, Q.W., Zhang, J.Y., Liu, D.S., Yi, Q.T., Chen, Y.C., Ma, H.H., Hu, L.M., 2020.
- 811 Nitrous oxide emissions from cascade hydropower reservoirs in the upper Mekong River.
 812 Water Res. 2020, 115582. https://doi.org/10.1016/j.watres.2020.115582
- 813 Sikar, E., Santos, M.A., Matvienko, B., Silva, M.B., Rocha, C.H.E.D., Santos, E., Bentes, A.P.B.,
- Rosa, L., 2005. Greenhouse gases and initial findings on the carbon circulation in two
 reservoirs and their watersheds. Verh. Internat. Verein. Limnol. 29, 573-576.
 https://doi.org/10.1080/03680770.2005.11902741
- 817 Straskraba, M., 1999. Retention Time as Key Variable of Reservoir Limnology. In Theoretical
- 818 Reservoir Ecology and its Applications; Tundisi, J.G., Straskraba, M., Eds.; Backhuys
- 819 Publishers: Leiden, pp. 385–410.

- Sturm, K., Yuan, Z., Gibbes, B., 2014. Methane and nitrous oxide sources and emissions in a
 subtropical freshwater reservoir, South East Queensland, Australia. Biogeosciences 11,
 5245–5258. https://doi.org/10.5194/bg-11-5245-2014
- Sun, Z.G., Wang, L.L., Tian, H.Q., Jiang, H.H., Mou, X.J., Sun, W.L., 2013. Fluxes of nitrous
 oxide and methane in different coastal Suaeda salsa marshes of the Yellow River estuary,
 China. Chemosphere 90, 856–865. https://doi.org/10.1016/j.chemosphere.2012.10.004
- Suntharalingam, P., and Sarmiento, J. (2000). Factors governing the oceanic nitrous oxide
 distribution: simulations with an ocean general circulation model. Global Biogeochem. Cy.
 14, 429–454. https://doi.org/10.1029/1999GB900032
- Syvitski, J.P.M., Vörösmarty, C.J., Kettner, A.J., Green, P., 2005. Impact of humans on the flux of
 terrestrial sediment to the global coastal ocean, Science, 308, 37–380.
 https://doi.org/10.1126/science.1109454.
- Taipale, S.J., Sonninen, E., 2009. The influence of preservation method and time on the δ¹³C value
 of dissolved inorganic carbon in water samples. Rapid Commun. Mass Sp. 23(16),
 2507-2510. https://doi.org/10.1002/rcm.4072
- Thornton, K.W., 1990. Sedimentary Processes. In: *Reservoir Limnology: Ecological Perspectives*;
 Thornton, K.W., Kimmel, B.L., Payne, F.E., Eds.; John Wiley & Sons: New York, pp. 43–70.
- Tian, L.L., Cai, Y.J., Akiyama, H., 2018. A review of indirect N₂O emission factors from
 agricultural nitrogen leaching and runoff to update of the default IPCC values. Environ.
 Pollut. 245, 300–306. https://doi.org/10.1016/j.envpol.2018.11.016
- Tian, L.L., Zhu, B., Akiyama, H., 2017. Seasonal variations in indirect N₂O emissions from an
 agricultural headwater ditch. Biol. Fertil. Soils 53, 651–662.
 https://doi.org/10.1007/s00374-017-1207-z
- 843 Turner, P.A., Griffis, T.J., Lee, X., Baker, J.M., Venterea, R.T., Wood, J.D., 2015. Indirect nitrous
- oxide emissions from streams within the US Corn Belt scale with stream order. Proc. Natl.
 Acad. Sci. U.S.A. 112, 9839–9843. https://doi.org/10.1073/pnas.1503598112
- Walter, S., Bange, H.W., Wallace, D.W., 2004. Nitrous oxide in the surface layer of the tropical
 North Atlantic Ocean along a west to east transect. Geophys. Res. Lett.31, 1–14.
 http://dx.doi.org/10.1029/2004GL019937
- 849 Wang, S.L., Liu, C.Q., Yeager, K.M., Wan, G.J., Li, J., Tao, F.X., Lü, Y.C., Liu, F., Fan, C.X.,
- 850 2009. The spatial distribution and emission of nitrous oxide (N_2O) in a large eutrophic lake in

- 851 eastern China: anthropogenic effects. Sci. Total Environ. 407(10), 3330–3337.
 852 https://doi.org/10.1016/j.scitotenv.2008.10.037.
- 853 Wang, X.F., He, Y.X., Yuan, X.Z., Chen, H., Peng, C.H., Yue, J.S., Zhang, Q.Y., Diao, Y.B., 854 Liu, S.S., 2017. Greenhouse gases concentrations and fluxes from subtropical small reservoirs 855 relation with watershed urbanization. Atmos. Environ. 154, 225-235. in http://dx.doi.org/10.1016/j.atmosenv.2017.01.047 856
- Wang, H.X., Zhang, L., Yao, X..L., Xue, B., Yan, W.J., 2017. Dissolved nitrous oxide and
 emission relating to denitrification across the Poyang Lake aquatic continuum. J. Environ. Sci.
- 859 52, 130-140. http://dx.doi.org/10.1016/j.jes.2016.03.021
- Wang, X.M., Hu, M.J., Ren, H.C., Li, J.B., Tong, C., Musenze, R.S., 2018. Seasonal variations of
 nitrous oxide fluxes and soil denitrification rates in subtropical freshwater and brackish tidal
 marshes of the Min River estuary. Sci. Total Environ. 616–617, 1404–1413.
 https://doi.org/10.1016/j.scitotenv.2017.10.175
- Wang, Z.Y., Xin, Y.Z., Gao, D.M., Li, F.M., Morgan, J., Xing, B.S., 2010. Microbial community
 characteristics in a degraded wetland of the Yellow River Delta. Pedosphere 20(4), 466–478.
 https://doi.org/10.1016/S1002-0160(10)60036-7
- Wanninkhof, R. 1992. Relationship between wind speed and gas exchange over the ocean. J.
 Geophys. Res. 97, 7373–7382. https://doi.org/10.1029/92jc00188
- Weiss, R.F., Price, B.A., 1980. Nitrous oxide solubility in water and seawater. Mar. Chem. 8(4),
 347–359. https://doi.org/10.1016/0304-4203(80)90024-9
- Welti, N., Hayes, M., Lockington, D., 2017. Seasonal nitrous oxide and methane emissions across
 a subtropical estuarine salinity gradient. Biogeochemistry 132, 55–69.
 https://doi.org/10.1007/s10533-016-0287-4
- Whitfield, C.J., Aherne, J., Baulch, H.M., 2011. Controls on greenhouse gas concentrations in
 polymictic headwater lakes in Ireland. Sci. Total Environ. 410: 217–225.
 https://doi.org/10.1016/j.scitotenv.2011.09.045
- Williams, C.J., Frost, P.C., Morales-Williams, A.M., Larson, J.H., Richardson, W.B., Chiandet,
 A.S., Xenopoulos, M.A., 2016. Human activities cause distinct dissolved organic matter
 composition across freshwater ecosystems. Glob. Change Biol. 22, 613-626.
 https://doi.org/10.1111/gcb.13094

- World Meteorological Organization, 2019. WMO Greenhouse Gas Bulletin No. 15 (25 November
 2019). https://library.wmo.int/doc_num.php?explnum_id=5455.pdf
- Wu, S., Chen, J., Li, C., Kong, D., Yu, K., Liu, S.W., Zou, J.W., 2018. Diel and seasonal nitrous
 oxide fluxes determined by floating chamber and gas transfer equation methods in agricultural
 irrigation watersheds in southeast China. Environ. Monit. Assess. 190, 122
 https://doi.org/10.1007/s10661-018-6502-0
- Xia, X.H., Zhang, S.B., Li, S.L., Zhang, L.W., Wang, G.Q., Zhang, L., Wang, J.F., Li, Z.H, 2018.
 The cycle of nitrogen in river systems: sources, transformation, and flux. Environ. Sci.:
 Processes Impacts 20, 863–891. https://doi.org/10.1039/C8EM00042E
- Xiao, Q.T., Xu, X.F., Zhang, M., Duan, H.T., Hu, Z.H., Wang, W., Xiao, W., Lee, X.H., 2019a.
 Coregulation of nitrous oxide emissions by nitrogen and temperature in China's third largest
- 892
 freshwater
 lake
 (Lake
 Taihu).
 Limnol.
 Oceanogr.
 64,
 1070–1086.

 893
 https://doi.org/10.1002/lno.11098

 </
- Xiao, Q.T., Hu, Z.H., Fu, C.S., Bian, H., Lee, X.H., Chen, S.T., Shang, D.Y., 2019b. Surface
 nitrous oxide concentrations and fluxes from water bodies of the agricultural watershed in
 Eastern China. Environ. Pollut. 251, 185–192. https://doi.org/10.1016/j.envpol.2019.04.076
- Xing, Y.P., Xie, P., Yang, H., Wu, A.P., Ni, L.Y., 2006. The change of gaseous carbon fluxes
 following the switch of dominant producers from macrophytes to algae in a shallow
 subtropical lake of China. Atmos. Environ. 40(40), 8034-8043
 https://doi.org/10.1016/j.atmosenv.2006.05.033
- Yan, F., Sillanpää, M., Kang, S., Aho, K.S., Qu, B., Wei, D., Li, X.F., Li, C.L., Raymond, P.A.,
 2018. Lakes on the Tibetan Plateau as conduits of greenhouse gases to the atmosphere. J.
 Geop hys. Res.-Biogeo. 123, 2091–2103. https://doi.org/10.1029/2017JG004379
- Yan, W.J., Yang, L.B., Wang, F., Wang, J.N., Ma, P., 2012. Riverine N₂O concentrations, exports
- to estuary and emissions to atmosphere from the Changjing River in response to increasing
 nitrogen loads. Global Biogeochem. Cy. 26, GB4006, https://doi.org/10.1029/2010GB003984
- Yang, H., 2014. China must continue the momentum of green law. Nature 509, 535-535.
 https://doi.org/10.1038/509535a
- Yang, H., Andersen, T., Dörsch, P., Tominaga, K., Thrane, J.-E., Hessen, D.O., 2015a.
 Greenhouse gas metabolism in Nordic boreal lakes. Biogeochemistry 126, 211-225.
 https://doi.org/10.1007/s10533-015-0154-8

- 912 Yang, H., Huang, X., Thompson, J.R., Flower, R.J., 2015b. Enforcement key to China's
 913 environment. Science 347(6224), 834-835. https://doi.org/10.1126/science.347.6224.834-d
- 914 Yang, H., Flower, R.J., 2012. Potentially massive greenhouse-gas sources in proposed tropical
 915 dams. Front. Ecol. Environ. 10(5), 234-235. https://doi.org/10.1890/12.WB.014
- Yang, H., Flower, R.J., Thompson, J.R., 2013. Sustaining China's water resources. Science
 339(6116), 141-141. https://doi.org/10.1126/science.339.6116.141-b
- 918 Yang, H., Xing, Y., Xie, P., Ni, L., Rong, K., 2008. Carbon source/sink function of a subtropical,
 919 eutrophic lake determined from an overall mass balance and a gas exchange and carbon burial
- 920 balance. Environ. Pollut. 151(3), 559-568. https://doi.org/10.1016/j.envpol.2007.04.006
- Yang, L.B., Yan, W.J., Ma, P., Wang, J.N., 2011. Seasonal and diurnal variations in N2O
 concentrations and fluxes from three eutrophic rivers in Southeast China. J. Geogr. Sci. 21(5),
 820–832. https://doi.org/10.1007/s11442-011-0882-1
- 924 Yang, P., Yang, H., Lai, D.Y.F., Guo, Q.Q., Zhang, Y.F., Tong, C., Xu, C.B., Li, X.F., 2020. 925 Large contribution of non-aquaculture period fluxes to the annual N₂O emissions from 926 J. Hydrol. 582, 124550. aquaculture ponds in Southeast China. 927 https://doi.org/10.1016/j.jhydrol.2020.124550
- Yoh, M., Terai, H., Saijo, Y., 1983. Accumulation of nitrous oxide in the oxygen deficient layer of
 freshwater lakes. Nature 301, 327–329. https://doi.org/10.1038/301327a0
- 930 Yu, J.H., Zhang, J.Y., Chen, Q.W., Yu, W.Y., Hu, L.M., Shi, W.Q., Zhong, J.C., Yan, W.X., 2018.
- 931 Dramatic source-sink transition of N₂O in the water level fluctuation zone of the Three Gorges
- 932 Reservoir during flooding-drying processes. Environ. Sci. Pollut. Res. 25, 20023-20031.
 933 https://doi.org/10.1007/s11356-018-2190-0
- Yu, Z.J., Deng, H.G., Wang, D.Q., Ye, M.W., Tan, Y.J., Li, Y.J., Chen, Z.L., Xu, S.Y., 2013. Nitrous
 oxide emissions in the Shanghai river network: implications for the effects of urban sewage
 and IPCC methodology. Global Change Biol. 19, 2999–3010.
 https://doi.org/10.1111/gcb.12290
- 938 Yu, Z.J., Wang, D.Q., Li, Y.J., Deng, H.G., Hu, B.B., Ye, M.W., Zhou, X.H., Da, L.J., Chen, Z.L.,
- 939 Xu, S.Y., 2017. Carbon dioxide and methane dynamics in a human-dominated lowland coastal
- 940 river network (Shanghai, China). J. Geophys. Res.-Biogeo. 122(7), 1738–1758.
- 941 https://doi.org/10.1002/2017JG003798

- Yu, Z., Yang, J., Amalfitano, S., Yu, X., Liu, L., 2015. Effects of water stratification and mixing on
 microbial community structure in a subtropical deep reservoir. Sci. Rep. 4, 5821.
 https://doi.org/10.1038/srep05821
- Zhang, L., Wang, L., Yin, K.D., Lü, Y., Zhang, D.R., Yang, Y.Q., Huang, X.P., 2013. Pore water
 nutrient characteristics and the fluxes across the sediment in the Pearl River estuary and
 adjacent waters, China. Estuar. Coast. Shelf S.133, 182-192.
 https://doi.org/10.1016/j.ecss.2013.08.028
- Zhao, Y., Wu, B.F., Zeng, Y., 2013. Spatial and temporal patterns of greenhouse gas emissions
 from Three Gorges Reservoir of China. Biogeosciences, 10(2), 1219–1230.
 https://doi.org/10.5194/bg-10-1219-2013
- Zhao, Y.Q., Xia, Y.Q., Kana, T.M., Wu, Y.C., Li, X.B., Yan, X.Y., 2013. Seasonal variation and
 controlling factors of anaerobic ammonium oxidation in freshwater river sediments in the
 Taihu Lake region of China. Chemosphere 93, 2124–2131.
 https://doi.org/10.1016/j.chemosphere.2013.07.063
- Zhou, Y.W., Xu, X.G., Han, R.M., Li, L., Feng,Y., Yeerken, S., Song, K., Wang, Q.L., Suspen
 ded particles potentially enhan ce nitrous oxide (N₂O) emissions in the oxic estuarine waters
 of eutrophic lakes: Field and experimental evidence. Environ. Pollut. 252, 1225–1234.
 https://doi.org/10.1016/j.envpol.2019.06.076
- 960 Zhu, D., Chen, H., Yuan, X.Z., Wu, N., Gao, Y.H., Wu, Y., Zhang, Y.M., Peng, C.H., Zhu, Q.A.,
- 961 Yang, G., Wu, J.H., 2013. Nitrous oxide emissions from the surface of the Three Gorges
- 962 Reservoir. Ecol. Eng. 60, 150–154. http://dx.doi.org/10.1016/j.ecoleng.2013.07.049
- 263 Zhang X.M., 2012. Spatiotemporal variations of nitrous oxide (N₂O) concentrations and flux in 17
- 964 reservoirs and their input rivers of Guangdong Province. Master Dissertation of Jinan
- 965 University (In Chinese with English Abstract).