

Hydrogen sulfide and environmental stresses

John T. Hancock

Department of Applied Sciences, University of the West of England, Bristol,
UK.

*Correspondence:

Prof. John T. Hancock

Faculty of Health and Applied Sciences,

University of the West of England, Bristol, BS16 1QY, UK.

john.hancock@uwe.ac.uk

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Abstract

Hydrogen sulfide (H₂S) is part of a suite of small reactive molecules which are known to be involved in cell signaling events in plants. It is produced by cells, can move around, including through membranes, and can be removed when no longer needed. It is perceived by cells, partly through persulfidation of proteins. Along with nitric oxide (NO) and reactive oxygen species (ROS) H₂S is involved in a range of stress responses, including following treatment with heavy metals, salt, temperature change and pathogen challenge. H₂S can lead to changes in the activity of antioxidants, cell signaling proteins such as mitogen activated protein kinases (MAPKs) and gene expression. Understanding how H₂S fits into cell signaling pathways may lead to advances in how treatment with H₂S or H₂S releasing donors may improve plant tolerance to stress, and hence plant growth and agricultural outputs.

Keywords: Antioxidants; cell signaling; drought; heavy metals; hydrogen peroxide; hydrogen sulfide; NaHS; nitric oxide; reactive oxygen species

Introduction

Despite its inherent toxicity hydrogen sulfide (H₂S) (Jiang *et al.*, 2016) has now become recognized as an important part of the suite of small diffusible compounds used by organisms in cell signaling (Hancock, 2017; Filipovic and Jovanovic, 2017). It has been known for some time that reactive compounds such as hydrogen peroxide (H₂O₂), along with other reactive oxygen species (ROS), can partake in signaling. In 1987 it was reported that endothelial derived relaxing factor in mammals was in fact the gas nitric oxide (NO) (Palmer *et al.*, 1987), and later the role of NO in plant signaling was recognized as being important (Neill *et al.*, 2002; Wilson *et al.*, 2008). Therefore, suggesting that other gases were also involved was not such an off-piste idea and so H₂S was being dubbed as the third gaseous transmitter (Wang, 2002) along with NO and carbon monoxide (CO), and it has been accepted that H₂S can act in cell signaling in plants (Wang, 2003; Hancock *et al.*, 2011; Wang 2012; Li, 2013; Lisjak *et al.*, 2013).

H₂S is inherently toxic (Jiang *et al.*, 2016). In *Escherichia coli* toxicity has been proposed to be through an oxidative damage mediated mechanism (Fu *et al.*, 2018) but in eukaryotes H₂S can act as a mitochondrial inhibitor, reducing the activity of Complex IV, lowering the electrochemical potential across the inner mitochondrial membrane (IMM) and reducing ATP generation (Dorman, 2002), although there is different sensitivity in different plants (Martin and Maricle, 2015). However, H₂S can also be produced by cells and can be used as a regulatory molecule (Filipovic and Jovanovic, 2017), impinging on cell signaling processes, especially those involving ROS and RNS. Interestingly, in mammals, at low concentrations H₂S can be used as mitochondrial reductant, and therefore an electron source for mitochondria (Bouillaud *et al.*, 2013; Módis *et al.*, 2016). Here electrons are fed into ubiquinone and hence

contribute to the generation of the IMM protomotive force and hence ATP production. From an evolutionary point of view perhaps this is not too surprising (Hancock, 2017). Organisms are known to use sulfur metabolism for energy conversion and early life would have been associated with sulfur rich thermal vents (Martin *et al.*, 2008). What is more surprising is the use of H₂S in cell signaling. It seems as though organisms have evolved to not only tolerate the presence of H₂S, but to use it as an integral part of a cell's control system (Hancock, 2017), in both animals and plants.

H₂S is small and therefore easy able to diffuse through cells, and being uncharged is likely to be able to traverse the hydrophobic nature of membranes: either the plasma membrane or those of organelles, not needing the assistance of proteins such as aquaporins (Mathai *et al.*, 2009). However, using erythrocyte membranes it has also be suggested that H₂S/HS⁻ movements across the lipid bilayer may involve exchange for Cl⁻ and use of the anion exchange protein AE1 in what is called the Jacobs-Stewart cycle (Jennings, 2013). Whether facilitated or not, movement of H₂S across membranes would facilitate its role in signaling, carrying the message from the point of production to the point of perception. The diffusion distance would however be determined by its removal, either enzymatic (Youssefian *et al.*, 1993; Tai and Cook, 2000) or by reacting with other cellular components.

Production of H₂S has been reported in cells. H₂S generating enzymes such as cystathione γ-lyase (CSE), cystathione β-synthase (CBS) and 3-mercaptopyruvate sulfurtransferase (3-MST) have been reported in animals (Prabhakar, 2012) but of more relevance here, desulfhydrases in plants (Alvarez *et al.*, 2010). DES1 in Arabidopsis has been shown to be a pyridoxal-5'-phosphate-dependent cysteine-degrading enzyme found in the cytosol (Alvarez *et al.*, 2010). In mitochondria, β-cyanoalanine synthase can also generate H₂S. The action of this

enzyme converts cysteine and cyanide to hydrogen sulfide and β -cyanoalanine (Hatzfeld *et al.*, 2000; Meyer *et al.*, 2003). The involvement of such enzymes in H₂S action is important to ascertain, as recently demonstrated by a study on seed germination, where in fact H₂S was concluded to have a marginal role (Baudouin *et al.*, 2016). Sulfide is also an intermediate in plant sulfur metabolism, being used along with O-acetylserine to generate cysteine (Calderwood and Kopriva, 2014), while bacteria to which plants may be interacting may also have the capacity to generate H₂S (Clarke, 1953). Therefore plants cells will most likely be exposed to sulfide, whether it is being used in signaling or not. Removal of H₂S can be catalyzed by enzymes such as O-acetylserine (thiol) lyase (Youssefian *et al.*, 1993; Tai and Cook, 2000), as well as by mitochondria in animals (Bouillaud *et al.*, 2013; Módis *et al.*, 2016), a type of activity which may also be relevant to plants. The oxidation of sulfide may also be important to the generation of further sulfur-based compounds which can also be involved in signaling, including persulfides, polysulfides and thiosulfate (Mishanina *et al.*, 2015), which may be important for stress responses.

The production, movement and removal of H₂S all make the role of this gas in cell signaling possible. Of course there will be issues of local production, use and removal so discussions of compartmentalization, as had been discussed for redox, ROS and RNS (Go and Jones, 2008; Noctor and Foyer, 2016), would be relevant to H₂S signaling, but such fine details have not been reported to date. New H₂S probes (Lin *et al.*, 2013; Chen *et al.*, 2018) in conjunction with confocal microscopy will no doubt shed light on such matters in the future. How different redox-related components are compartmentalized, either in the same manner or differently, will determine the type of competition there will be between them and therefore the ultimate signaling response.

It may be worth at this point noting that experimentally H₂S is often supplied to a system under study using compounds which release H₂S into solution, rather than the use of the gas *per se*. The kinetics of release varies with different donors. Compounds such as NaHS tend to release H₂S very quickly, while newer donors have slower release kinetics (such as GYY3147), or are targeted to specific organelles (such as AP39). For a review of H₂S donors see Powell *et al.* (2018).

Intracellular effects of H₂S

As well as being able to inhibit mitochondrial activity (Dorman, 2002; Martin and Maricle, 2015) H₂S may also be used by mitochondria (Bouillaud *et al.*, 2013) as an electron donor. In fact, the mechanisms of how H₂S may interact and regulate mitochondrial activity is quite complex, and may involve feeding electrons through sulfide:quinone oxidoreductase (SQR) and Complex II, interacting with free radicals, and controlling cell signaling pathways (Módis *et al.*, 2016). However, for H₂S to be useful in cell signaling and stress responses H₂S needs to be perceived and its presence acted on. Normally such activity is triggered by a ligand (in this case H₂S) arriving at a receptor and initiating a signal transduction pathway. It has been suggested that H₂S is too small to dock to a receptor in the classical sense (Moore and Whiteman, 2015), but in animals the protein VEGFR2 has been suggested as a candidate as it can react directly with H₂S. It is likely that in plants a similar chemistry is involved in H₂S perception (as discussed below).

One of the direct influences of H₂S will be on the presence of the other stress signals, such as ROS and NO. H₂S can react with peroxynitrite (ONOO⁻) for example (Carbellal *et al.*, 2011), so removing the latter from solution, and so removing its signaling capacity. NO can also react with H₂S. This has two consequences. Firstly

the presence of both H₂S and NO will be reduced, and so removing the ability of both to cause further signaling. Secondly a new compound is produced, nitrosothiol, which itself may have a signaling property (Whiteman *et al.*, 2006). The chemistry of H₂S was extensively explored by Li and Lancaster (2013): it will not be stable and unreactive in cells, which would limit its capacity to move and act as a distant signal. Here the notion of compartmentalization is needed to be considered once more.

A major target for many reactive molecules in signaling is the thiol group on proteins. The cysteine –SH group can undergo attack (often after removal of the proton) in a variety of ways (Hancock, 2009). It can be oxidized by H₂O₂ to the sulfenic acid group. This modification is akin to phosphorylation, in that the thiol can be toggled between two states: modified and un-modified. Therefore the activity of a protein can be toggled between two states by the action of H₂O₂, for example between active and inactive. With increased concentrations of H₂O₂ the thiol group may be further oxidized to the sulfinic acid and further more to the sulfonic acid, the latter two probably being irreversible changes. On the other hand NO can modify the thiol by S-nitrosation (often referred to as S-nitrosylation), which can be identified using the biotin switch assay (Jaffrey and Snyder, 2001; Grennan, 2007). Again, this is akin to phosphorylation, giving two states between which the protein may toggle. Other modifications include glutathionylation (Sun *et al.*, 2013), especially important when you consider the high concentration of glutathione in cells (Schafer and Buettner, 2001). In a similar manner H₂S can lead to the modification of the thiol group, by a process known as S-sulfhydration (or persulfidation) (Romero *et al.*, 2011; Paul and Snyder, 2015; Aroca *et al.*, 2017a). In each of these cases the modification of the thiol is different and there is no reason to assume that the final activity of the protein would be the same (Figure 1). Clearly, the proportion of the

protein which has been modified in each way will depend on the local concentrations of the reactive molecules, which can be altered by each other: that is, H₂S can remove NO, as can superoxide anions etc. Local concentrations will also be determined by compartmentalization. In plants persulfidation has been shown to be involved in the regulation of the Krebs's cycle, glycolysis and the Calvin cycle, showing that it has a central role in regulating metabolic processes (Aroca *et al.*, 2017a), and not just a trivial chemical anomaly. However, further work is needed here to fully understand the role of H₂S and persulfidation in controlling metabolic processes in plants.

A good example of a protein that is covalently modified by a range of small reactive molecules is glyceraldehyde 3-phosphate dehydrogenase (GAPDH) (Holtgrede *et al.*, 2008). This protein is central to glycolysis but isoforms are also found in the chloroplasts of cells (Martin and Cerff, 2017). GAPDH can be oxidized and S-nitrosated (Hancock *et al.*, 2005; Lindermayr *et al.*, 2005), and can be modified by H₂S (Aroca *et al.*, 2015; Aroca *et al.*, 2017b). On modification this protein can move from the cytoplasm, where it is involved in glycolysis, to the nucleus where it can control transcription factors and hence gene expression. Therefore, thiol modification has a profound effect on the protein's function, bestowing on it a completely disparate activity.

Immensely important targets for covalent modification by reactive signals are transcription factors themselves. As well as often being phosphorylated, a process which can be enhanced by the presence of reactive signals – as discussed further below – transcription factors can be directly modified. Albeit in animals, a good example is NF-κB which can be modified by a range of mechanisms (Perkins, 2006),

including by NO and H₂S (Sen *et al.*, 2012). Certainly H₂S has been shown to have large effects on the gene expression profiles of plants cells (Li *et al.*, 2017b).

One of the influences of H₂S is that it can increase the levels of glutathione (De Kok *et al.*, 1985). Glutathione is found in high concentrations in cells and has an immensely important role in maintaining the intracellular redox balance, and perhaps preventing cells from entering programmed cell death (apoptosis) or necrosis (Schafer and Buettner, 2001). More recently in a study on water stress in wheat seedlings following NaHS treatment there were effects on both glutathione and ascorbate metabolism (Shan *et al.*, 2011), with levels of reduced glutathione and reduced ascorbate both being increased. Again, along with soluble thiols such as glutathione, ascorbate is important to control cellular redox (Noctor *et al.*, 2018). If intracellular redox becomes oxidized levels of important reduced molecules such as NADH/NADPH are harder to maintain and apoptosis (programmed cell death) or necrosis can be initiated (Schafer and Buettner, 2001). Therefore, the presence of H₂S may help to maintain the GSH. Loss of glutathione would alter the redox status of the plant cell (Foyer *et al.*, 2001) as the concentrations of oxidised and reduced glutathione feed into a squared relationship in the Nernst equation (Schafer and Buettner, 2001), meaning that the total glutathione present (i.e. GSH + GSSG) helps to dictate the overall redox state of the cell, not just the GSH/GSSG ratio.

Concentration levels of reduced glutathione may decrease during oxidative stress, when ROS accumulation increases, and hence, by increasing intracellular glutathione, H₂S can have an important influence on cell function, particularly in times of stress. The importance of changes in antioxidants is highlighted, perhaps, by the work on postharvest. H₂S has been shown to increase the shelf life of strawberries, an effect mediated by antioxidant levels (Hu *et al.*, 2012). Similar

studies and effects have also been reported for other fruit such as kiwifruit (Zhu *et al.* 2014) and grape (Ni *et al.*, 2015), although effects of ethylene signaling have also been reported (Li *et al.*, 2017a). Therefore, manipulation of H₂S may alter intracellular redox but may also be of commercial importance too.

In a similar manner, gaseous signal molecules such as H₂S can influence proline levels (He and He, 2018) which is immensely important for the maintenance of redox and protecting against oxidative stress, amongst other things (Kavi Kishor and Sreenivasulu, 2014). As an example of the importance of such redox modulators, Wang *et al.* (2012) suggested that the effects of H₂S on salinity tolerance were through effects on oxidative stress, and possibly NO. Therefore, overall the influence of H₂S on the intracellular redox status of cells is an important issue to be aware of.

Stress responses involving H₂S

Stress responses in cells often involve reactive signaling molecules. It has long been known that plant stress leads to an increase in ROS accumulation (Mittler, 2002; Baxter *et al.*, 2014). In response to a wide range of stresses, such as heat, light, heavy metals and pathogen challenge plant cells will accumulate H₂O₂ which will lead to a range of downstream effects, mediated by signal transduction components such as mitogen activated protein kinases (MAPKs), and resulting in altered gene expression. In a similar way, NO is also integrally involved in plant stress responses (Arasimowicz and Floryszak-Wieczorek, 2007), again being accumulated in response to similar initiators and resulting in similar cellular alterations. Therefore, the involvement of other small gaseous signaling molecules such as H₂ (Cui *et al.*, 2014) and H₂S would be expected. Previous reviews have discussed how H₂S may

mediate plant responses, as shown in Figure 2, and how there may be an interplay with other signaling molecules (Hancock and Whiteman, 2014; 2015), as illustrated in Figure 3. Below is a short discussion on some of the current reports of H₂S mediating plant stress responses.

Being sessile plants have to tolerate the temperature that their environment dictates and this is often not optimal (Bita and Gerats, 2013). H₂S has been implicated in the tolerance to both high and low temperatures by several research groups. Stuiver *et al.*, (1992) investigated freeze tolerance in wheat seedlings and found that H₂S altered the cellular levels of amino acids and sugars. They also measured water-soluble sulfhydryl content (discussed in Hancock and Whiteman, 2018), which was primarily glutathione and found that this too increased following H₂S fumigation. At the other end of the temperature scale, heat tolerance is also improved by the treatment with H₂S or donors. It was found that H₂S – in the form of NaHS – increased germination of maize seeds under heat stress, and further enhanced tissue viability and lowered malondialdehyde (MDA) accumulation caused by heat treatment (Li *et al.*, 2013a). It was concluded that H₂S action was being mediated by an increase in proline levels in cells. Subsequently, NaHS, acting as a H₂S donor, enhanced salicylic acid (SA)-induced heat tolerance in maize (Li *et al.*, 2015), an effect blocked if a H₂S biosynthesis inhibitor or H₂S scavenger was used. The authors suggested that H₂S was acting downstream of SA. In tobacco, the H₂S donor NaHS was used to increase the survival of cell cultures under heat stress (Li *et al.*, 2012b). Here it was suggested that the effects were mediated by the entry of extracellular calcium ions across the plasma membrane and that intracellularly the effects were reliant on the action of calmodulin, a ubiquitous calcium binding protein. In another study, strawberry roots (*Fragaria x ananassa* cv. 'Camarosa') were

exposed to an acute heat shock in the presence and absence of NaHS. MDA, H₂O₂ and NO were all lower in H₂S treated tissues. Ascorbate and glutathione metabolism were suggested to be involved in preserving tissues against heat-induced damage and gene expression of several antioxidants and heat shock proteins was induced, including catalase, superoxide dismutase, HSP70, HSP80 and HSP90. Aquaporin was also induced (Christou *et al.*, 2014). More recently, the foliar application of NaHS on wheat seedlings showed an increase in heat tolerance. As would be expected, antioxidant levels were increased, signs of oxidative stress, such as MDA, were decreased and the authors suggested that foliar application of H₂S donors may be beneficial (Zhang *et al.*, 2016).

Another environmental stress often needing to be tolerated by plants is the presence of heavy metals (Nikalje and Suprasanna, 2018). Previously the role of H₂S in plants in response to cadmium, aluminum, chromium, copper and lead have been discussed (Hancock and Whiteman, 2015). Li *et al.* (2012c) suggested that NO was involved in the alleviation of cadmium stress, while with chromium stress NaHS promoted plant growth and photosynthesis (Zhang *et al.*, 2010a; Ali *et al.*, 2013). In wheat, NaHS could relieve the reduction of germination of seed caused by copper stress (Zhang *et al.*, 2008), while antioxidant levels were increased and MDA and H₂O₂ accumulation was reduced. Lead stress in oilseed rape (*Brassica napus* L) was alleviated by NaHS, with improved growth, photosynthesis and cell structure (Ali *et al.*, 2014). Finally, in aluminum stress NaHS increased the activities of antioxidant enzymes and citrate secretion. The expression of the citrate transporter gene was enhanced as was that of the PM H⁺-ATPase (Chen *et al.*, 2012), while H₂S alleviated the toxicity of Al to germinating wheat seedlings (Zhang *et al.*, 2010b).

The role in heavy metal stress by H₂S has also recently reviewed by others (He *et al.*, 2018), and such research continues. Recently chromium treatment of maize (*Zea mays* L.) led to oxidative stress symptoms in some tissues, but not all. However, the effects were alleviated by the addition of NO and H₂S donors (Kharbech *et al.*, 2017). ROS was found to be important in the H₂S effects seen on Zn treatment of pepper (*Capsicum annuum* L.). Exogenous NaHS enhanced plant growth and fruit yield, altered levels of antioxidants and suppressed MDA and H₂O₂ content (Kaya *et al.*, 2018). In a study on roots, Lv *et al.* (2017) showed that 4μM Cd led to an increase in H₂S at the root tip, an effect blocked by a H₂S scavenger or an inhibitor of H₂S biosynthesis. It was concluded that part of the action of H₂S was through the modulation of the H₂O₂ and O₂^{·-} levels at the tip. Also recently, H₂S was shown to alleviate aluminum toxicity in rice (Zhu *et al.*, 2018). Al content in the root tips was reduced after treatment with NaHS. NaHS pretreatment lowered the negative charge in cell walls and masked Al binding sites by upregulation of OsSTAR1 and OsSTAR2 in roots. The gene, OsFRDL4, which is involved in citrate exudation was over-expressed. Intracellular Al was altered, as there was an increase in the translocation of Al to the vacuole: OsALS1 was over-expressed. MDA and H₂O₂ content was decreased following changes in antioxidant levels. The authors also pointed out that H₂S exhibited crosstalk with NO signaling, emphasizing that H₂S was not working alone, as shown in Figure 3.

This latter point highlights the importance of considering the reactive molecules together (Hancock and Whiteman, 2014), as well as interactions with other signals. H₂S toxicity itself appears in some instances to require ROS and NO. The inhibition of growth caused by H₂S was reported to be auxin dependent but a further study showed that MAPKs were involved, specifically MPK6, but also was

305 accompanied by an increase in ROS accumulation and NO production (Zhang *et al.*,
306 2017). In rice (*Zizania palustris*) sulfide toxicity was alleviated by the presence of iron
307 (Fort *et al.*, 2017), itself a metal often involved in ROS metabolism (Koskenkorva-
308 Frank *et al.*, 2013).

309 The control of water use in plants is extremely important. Plants may
310 experience too much or too little water, a topic of increasing importance with the
311 onset of global climate change (Jasechko, 2018). Under waterlogging stress the
312 application of NaHS up to 0.1mM alleviated the symptoms of the stress and it was
313 suggested that this was partly mediated by a reduction in ROS accumulation (Wei *et al.*,
314 2017a). In a similar study of submerged macrophytes exposed to hypoxia and
315 H₂S, in *Potamogeton crispus* and *Myriophyllum spicatum* there was an increase in
316 oxidative stress, increased H₂O₂ and MDA when exposed to sulfide in the form of
317 NaHS. However, this was not replicated in other species such as *Egeria densa* and
318 *Potamogeton oxyphyllus* (Parveen *et al.*, 2017).

319 Plants often are exposed to conditions where water is less than optimal.
320 Drought stress has been studied for many years and the role of H₂S on the control of
321 stomatal closure has been of some debate. H₂S may cause stomatal opening in some
322 instances but closure under others (Garcia-Mata and Lamattina, 2010; 2013; Lisjak *et al.*,
323 2010; 2011). More recently, this apparent anomaly has been further investigated
324 (Honda *et al.*, 2015). They showed that short-term exposure to a H₂S donor caused
325 closure, but that prolonged exposure to the H₂S donor caused increased stomatal
326 apertures. Of particular significance, they showed that the effects of H₂S were
327 mediated by 8-mercapto-cGMP. Downstream of this signaling molecule it was found
328 that Ca²⁺, cADP ribose and slow anion channel 1 were involved. It has also been
329 suggested that H₂S interacts with abscisic acid (ABA) to have effects (Jin *et al.*, 2013)

and that H₂S is involved in ethylene signaling which leads to stomatal closure (Liu *et al.*, 2011). Using a DES1 mutant in Arabidopsis the role of H₂S and its interactions with NO and ABA were further investigated (Scuffi *et al.*, 2014). It was found that ABA induced DES1 expression, and that DES1 was needed for ABA-dependent NO generation, with NO being downstream of H₂S in the signaling pathway. The authors concluded that DES1, and hence H₂S accumulation, should be included as a component of the ABA pathway. The involvement of ABA has been studied further recently, showing cross-talk between ABA signaling and H₂S during drought responses (Ma *et al.*, 2016). The expression levels of genes involved in ABA metabolism were altered, somewhat differently in roots and leaves, although expression levels of ABA receptors were increased in both roots and leaves following H₂S treatment during drought stress. More recently drought-responsive genes which may be regulated by H₂S were studied in wheat (*Triticum aestivum* L.). Over 7500 genes were identified as being worth further investigation (Li *et al.*, 2017b). Genes involved in iron transport were of significance as were pathways for protein processing in the endoplasmic reticulum and fatty acid degradation. H₂S was also shown to influence plant hormone signaling pathways, including transcription factors and protein kinases. Such studies highlight the wide-ranging influences of H₂S on plant growth and survival.

Also looking at gene expression Wei *et al.* (2017b), suggested that H₂S positively enhances the expression of senescence associated genes (SAGs). In their system H₂S suppressed chlorophyll degradation of detached leaves and they suggested that S-nitrosoglutathione reductase was important, implicating NO metabolism in the mechanism.

H₂S alleviated the effects of salt stress of roots in *Arabidopsis*. Using NaHS ion transport was shown to be important, but there was also a dependence on H₂O₂ (Li *et al.*, 2013b). It appeared that H₂S increased intracellular H₂O₂ by regulating the activities of two enzymes: glucose-6-phosphate dehydrogenase (G6PDH) and plasma membrane (PM) NADPH oxidase. This seems to be the wrong way around, as this would increase oxidative stress in the tissues unless the increases are carefully controlled by antioxidants and/or compartmentalisation. In contrast, H₂S was also found to alleviate oxidative stress in sweet potato undergoing osmotic stress, induced with PEG-6000 (Zhang *et al.*, 2009b). NaHS treatment alleviated chlorophyll loss, while antioxidants such as catalase and superoxide dismutase (SOD) were increased. Both H₂O₂ and MDA were decreased on NaHS treatment. In a subsequent study NaHS was found to increase wheat seed germination during osmotic stress with PEG-6000 (Zhang *et al.*, 2010c). Again, H₂O₂ and MDA were reduced, while catalase and ascorbate peroxidase activities were increased. Interestingly, SOD was not affected in this study.

Both ROS and NO have been extensively studied in pathogen challenge of plants (Bellin *et al.*, 2013), but less is known about the involvement of H₂S. Some work has been reported with fungi infections (Bloem *et al.*, 2012) while effects of H₂S on plant pathogenic bacteria have been studied (Neale *et al.*, 2017).

Other volatile compounds need to be considered too (Piechulla *et al.*, 2017) and interestingly H₂S was shown to be downstream of methane in the induction of adventitious root development in cucumber (Kou *et al.*, 2018). One gas that is gaining prominence in the literature due to its ability to ameliorate disease symptoms and to alleviate stress is H₂ gas (Cui *et al.*, 2014; Wilson *et al.*, 2018). How cell signaling by this gas intermeshes with that of H₂S needs to be determined in the future.

379

380 **Fitting H₂S into signaling pathways**

381 The placement of H₂S in signaling pathways is important: is it upstream or
382 downstream of other effectors? In a study with tomato and high salt stress, DaSilva *et*
383 *al.* (2018) concluded that H₂S was downstream of NO, mitigating oxidative stress and
384 helping the plant to tolerance the stress it was exposed to. Others have also suggested
385 that H₂S was downstream to NO (Li *et al.*, 2013c) and also downstream to salicylic
386 acid (Li *et al.*, 2015). In banana H₂S alleviated post-harvest ripening and senescence
387 by a reduction of oxidative stress but also an inhibition of ethylene signaling (Ge *et al.*,
388 2017). The wider issue of the involvement of sulfur-based compounds on
389 phytohormones was reviewed elsewhere (Hasanuzzaman *et al.*, 2018). Also in tomato
390 H₂S was shown to be upstream of NADPH oxidase (RBOH1) and H₂O₂ accumulation
391 (Mei *et al.*, 2017). In *Arabidopsis* roots H₂S was also shown to regulate NADPH
392 oxidase, as well as glucose-6-phosphate dehydrogenase (G6PDH) (Li *et al.*, 2013b).
393 In guard cells of *Arabidopsis* H₂S was shown to regulate NADPH oxidase activity, and
394 hence ROS accumulation, while at the same time increased phospholipase D δ -derived
395 phosphatidic acid levels, and so alter further signaling pathways in the cells (Scuffi *et*
396 *al.*, 2018).

397 MAPKs have been shown to be involved in some studies. In discussion
398 above MAPK was implicated in the toxicity response to H₂S, but in a study on cold
399 stress in *Arabidopsis thaliana* MPK4 was shown to be important. H₂S inhibited
400 stomatal opening under cold stress, and it was concluded that H₂S was upstream of
401 the MAPK pathway (Du *et al.*, 2017).

402 It has been suggested that H₂S has a modulating effect on ROS and RNS
403 metabolism in some cases (Hancock and Whiteman, 2014). The over-accumulation of

ROS, or RNS, can lead to detrimental effects on cells and tissues, perhaps triggering programmed cell death. Such effects of ROS and RNS may be mediated by their influence on the intracellular redox poise of the cell (Schafer and Buettner, 2001). As H₂S can directly react with some ROS species and NO then the accumulation of these signals may be lowered. Further, H₂S can react with enzymes which generate ROS and RNS, and can influence the levels of antioxidants which lower ROS and RNS, and so restore the cellular redox poise. This has been previously discussed in more detail (Hancock and Whiteman, 2014; 2015), but such an influence of H₂S would account for many of the results of H₂S during cellular stress. Further, it has been argued recently that the maintenance of the cellular redox, perhaps with H₂S's influence, is essential to allow the correct functioning of redox-based signals such as ROS and RNS (Hancock and Whiteman, 2018). It is undoubtedly a fine balance between the levels of ROS, RNS, H₂S, antioxidants and redox poise that will allow a cell to mount the correct response to any stress put upon it.

Conclusions and future perspectives

H₂S has been implicated in a large range of plant cell functions, from germination (Zhang *et al.*, 2008; Li *et al.*, 2012a; Dooley *et al.*, 2013), root development (Zhang *et al.*, 2009a; Lin *et al.*, 2012), stomatal aperture control (Garcia-Mata and Lamattina, 2010; Lisjak *et al.*, 2010), to flower senescence (Zhang *et al.*, 2011). It has also been shown to be involved in a myriad of stress responses, including heavy metal stress, freezing, heat, salt stress and oxidative stress.

It is known that H₂S has an important role in plant growth and stress responses but it is becoming clear that numerous volatile compounds may need to be considered (Piechulla *et al.*, 2017), including H₂S and H₂ gas (Cui *et al.*, 2014). It

is a holistic approach to the effects of such compounds that is needed to fully understand how plants can respond to their environment. Many redox-based compounds are clearly involved in the control of cellular function and it is the interactions between them that needs to be considered. Much work over many years has concentrated on ROS and RNS metabolism in plants, but the influence of H₂S needs to be understood in different tissues under different conditions, including a wide range of stresses. Furthermore, the effects on intracellular redox and thiol-controlled proteins are vital to understand so that redox-based metabolism, be it that of ROS, RNS or H₂S can be modulated to aid plant growth under stressful conditions. This is perhaps even more important now that world climate change is being perceived to be having tangible effects (Makuvaro *et al.*, 2018).

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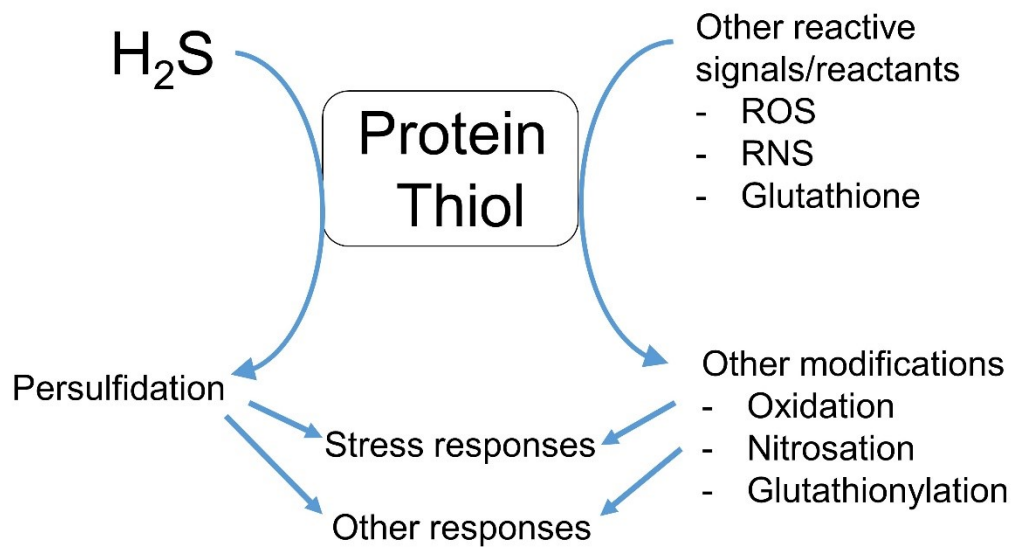
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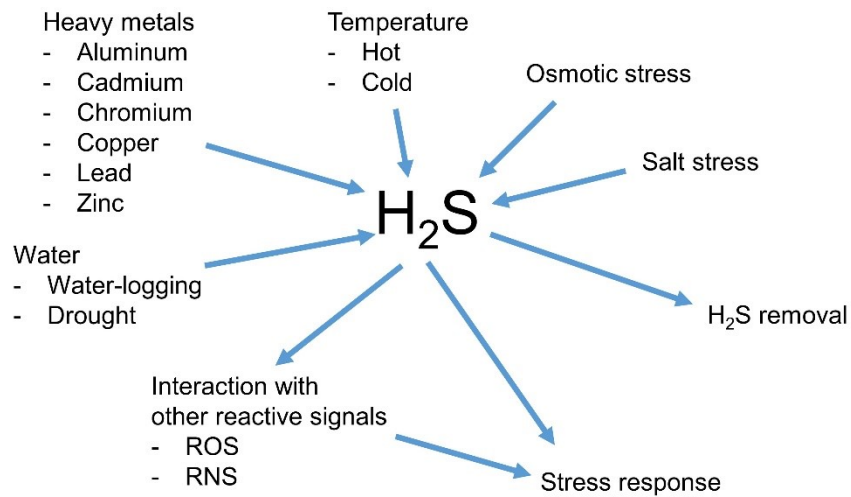
Figure Legends

Figure 1: Hydrogen sulfide may directly modify protein thiol groups.

The thiol group (-SH) on proteins may be modified by H₂S but thiol groups can be reacted with other reactive signaling compounds in the cell. Therefore, a competition may be set up in the cell, with the end result dependent on the prominence of the signaling molecules present.



839 Figure 2: Hydrogen sulfide is a central component in plant stress responses.
840 Many stresses may lead to the accumulation of H₂S in plants and this may lead to
841 the alleviation of cell stress.



842

843

Figure 3: Downstream effects of H₂S that may lead to a response.

There are many ways in which H₂S may influence cellular function: interaction with other reactive signals; modulation of enzyme activities; persulfidation; effects on antioxidants. These are not exclusive and a combination of downstream events may lead to the ultimate response.

